

New Therapeutic Targets for Osteoarthritis Pain

SLAS Discovery
2017, Vol. 22(8) 931–949
© 2017 Society for Laboratory
Automation and Screening
DOI: 10.1177/2472555217716912
journals.sagepub.com/home/jbx


David A. Walsh^{1,2,3,4,5} and Joanne Stocks^{1,2,3}

Abstract

Osteoarthritis (OA), the most common form of arthritis, causes pain and disability, as well as emotional distress. While total joint replacement is one of the most effective treatments available for improving the quality of life in people with severe OA, it is not suitable for all patients and all joints. Current pharmacological analgesics have limited efficacy, and their use is often restricted by adverse events. Medications that might reduce pain by slowing or preventing structural disease remain elusive. Our increasing understanding of the complex mechanisms that underlie OA pain offers a wide range of potential new treatment targets. New drugs for OA pain might come from repurposing those developed for other conditions, as well as novel compounds targeting pain mechanisms specific to the joint. Here we discuss the mechanisms of OA pain and its therapeutic implications. We explore evolving treatment modalities, including combination treatment. We review recent research and patents pointing to future OA therapies. We discuss the potential for biomarkers to facilitate drug development and targeting.

Keywords

osteoarthritis, pain, inflammation, osteoclasts, peripheral sensitization, central sensitization

What Is Osteoarthritis?

Osteoarthritis (OA) is the most common disease of synovial joints, and a major source of pain, disability, and distress. Peak OA incidence is from the sixth decade, and with increasing longevity and employment into older age, OA has become an increasing burden on the economies and healthcare systems of the developed world. In the absence of a cure, OA increases in prevalence with increasing age, such that everyone who survives beyond the age of 70 might expect to develop OA in at least one joint. However, far from being simply an inevitable consequence of aging, OA might be better viewed as a heterogeneous group of related diagnoses, each displaying discrete genetic associations and precipitating and aggravating factors. For a majority of those unfortunate enough to experience end-stage disease, surgical arthroplasty might offer substantial pain relief and improve quality of life. However, arthroplasty is not currently possible for all joints, nor suitable or successful for all patients. The mainstays of treatment for most people with OA comprise analgesia and exercise. Existing drugs often offer little benefit, or risk important adverse events. Medications that might slow or prevent structural disease remain elusive.

OA has been defined both clinically and radiographically. Pain and crepitus are sufficient to make a clinical classification of knee OA in those aged over 40 years.¹ Radiographic characteristics evidence new bone formation (osteophytosis and subchondral sclerosis), cartilage loss

(joint space narrowing), and subchondral cysts. Of these, osteophytes are most specific for OA, whereas joint space narrowing also occurs in other diseases, including rheumatoid arthritis (RA). However, the relationship between symptoms and radiographic appearance is only weak. Patients with advanced radiographic changes might be asymptomatic, whereas knee pain is common before radiographic features are apparent. Knee pain is predictive of subsequent radiographic OA development,² indicating a preradiographic stage in some patients, whereas for others, radiographic change precedes the onset of pain.

Attempts to define “early” OA have focused on the distinction from end-stage disease, in an attempt to identify subgroups of people for whom treatments that facilitate

¹Arthritis Research UK Pain Centre, University of Nottingham, UK

²Division of Rheumatology, Orthopaedics and Dermatology, School of Medicine, University of Nottingham, UK

³NIHR Nottingham BRC, UK

⁴Sherwood Forest Hospitals NHS Foundation Trust, UK

⁵Nottingham University Hospitals NHS Trust, UK

Received Jan 15, 2017, and in revised form April 26, 2017. Accepted for publication May 16, 2017.

Supplementary material is available online with this article.

Corresponding Author:

David A. Walsh, Arthritis Research UK Pain Centre, Academic Rheumatology, University of Nottingham, Clinical Sciences Building, City Hospital, Nottingham, NG5 1PB, UK.
Email: David.walsh@nottingham.ac.uk

joint repair might offer secondary prevention. Tissue engineering approaches, for example, might eventually reduce disease burden and need for arthroplasty. However, the boundary between early OA and normality is not as easy to define.³ Histopathological features, such as cartilage fibrillation and proteoglycan loss, might be seen in young adults, for example, those with chondromalacia patellae, but need not necessarily progress to OA in later life.

OA subgroups have been traditionally defined according to joint distribution (hip, knee, spine, and generalized nodal OA) or by predisposing factors (primary or secondary to injury or other joint disease). The impact of OA differs between different anatomical sites, with OA of large weight-bearing joints (hip, knee, and less often, ankle) often causing major disability, and OA of small joints of the hands impairing grip and fine manipulation.

Secondary OA offers potential for preventative treatments. RA in its early stages causes pain through immune synovitis, whereas secondary OA in later disease leads to persistent pain even after inflammation is fully suppressed. Immune modulation in early RA reduces joint damage as well as pain.⁴ Optimal management of joint injuries focuses on the prompt relief of pain and functional restoration, and further research is required to optimize interventions that will prevent subsequent OA development. Reconstructive surgery for meniscal or cruciate ligament injuries has displayed limited capacity to retard the onset of posttraumatic OA, and current research focuses on the potential for tissue engineering approaches or medical interventions to improve long-term prognosis.⁵

The contribution of OA to back pain remains controversial. Structural associations of low back pain remain even more difficult to define than in knee OA, with potential origins of pain in discs, as well as facet joints and periarticular structures.^{6,7} Radiofrequency denervation procedures designed to reduce nociceptive signals from facet joints have increasing evidence of benefit in carefully selected cases with low back pain,⁷ although radiographic facet OA is not predictive of pain outcomes.⁶

Mechanisms of OA Pain: Therapeutic Implications

Biomechanical Model of OA Pain

Although OA has been recognized since ancient times, the mechanisms by which OA pathology causes pain have only recently been elucidated. Early perspectives viewed OA as primarily a mechanical condition of wear and tear, based on pathological cartilage loss and typical pain that is exacerbated by weight bearing and movement. Low muscle strength predicts worsening OA pain,⁸ and muscle strengthening exercises can reduce OA pain.⁹ This biomechanical model has led to treatments that offer relief to many, including exercise regimens, orthotics, and arthroplasty.

Pain improvement follows joint replacement surgery, although its precise mode of action remains uncertain. Arthroplasty removes diseased cartilage and subchondral bone, and reinstates a biomechanical and chemical barrier between synovial cavity and bone marrow spaces. Subchondral nerves are sectioned. By contrast, the synovium and periarticular structures remain intact or regenerate following arthroplasty. Synovitis might persist, but the foreign body reactions found postarthroplasty differ from the cellular and biochemical characteristics of the OA joint.¹⁰ An unfortunate and important minority of people continue to experience significant pain after arthroplasty (estimated as 5% after total hip arthroplasty and up to 20% after knee replacement surgery), indicating either new pathological mechanisms consequent to surgery and prosthesis, or persistent OA pain mechanisms that are not resolved by arthroplasty.

Structural Associations of OA Pain

The biomechanical model led to extensive research being translated into clinical practice using imaging as a biomarker of OA. Radiographic joint space narrowing (**Fig. 1a,b**) has been more consistently associated with OA pain than has osteophytosis, and the relative contributions of new bone formation to OA pain or to articular repair remain uncertain. Treatments should aim to selectively restore or preserve structures in which pathology causes symptoms, while retaining capacity for repair. Knee joint space narrowing on plain weight-bearing radiographs, classically affecting the medial tibiofemoral compartment, is suggestive of articular cartilage thinning, but might also be caused by meniscal extrusion. Varus instability can increase lateral radiographic joint space despite significant chondropathy. Radiographs therefore provide only limited information about articular health, and associations between radiographic findings and pain tend to be weak.

Magnetic resonance imaging (MRI) has offered a more detailed assessment of the structural OA changes that might be associated with (and therefore mediate) pain (**Fig. 1c,d**). MRI permits visualization of the articular cartilage, and therefore accurate assessment of cartilage depth and focal defects. Meniscal location and tears can be evaluated. Synovitis might be indicated by synovial hypertrophy, by gadolinium enhancement by increased blood flow, or by effusion. MRI reveals lesions in subchondral bone (bone marrow lesions [BMLs]). These were originally considered to represent edema but are now recognized as heterogenous pathologies associated with fibrovascular replacement of bone marrow, microfracture, or increased bone turnover.¹¹ Vascular penetration from subchondral bone into the normally avascular noncalcified articular cartilage¹² and overlying cartilage defects¹¹ often accompany this subchondral pathology. Each OA structural characteristic is correlated with other morphological changes, as expected with diverse aspects of a single disease. Each is associated with

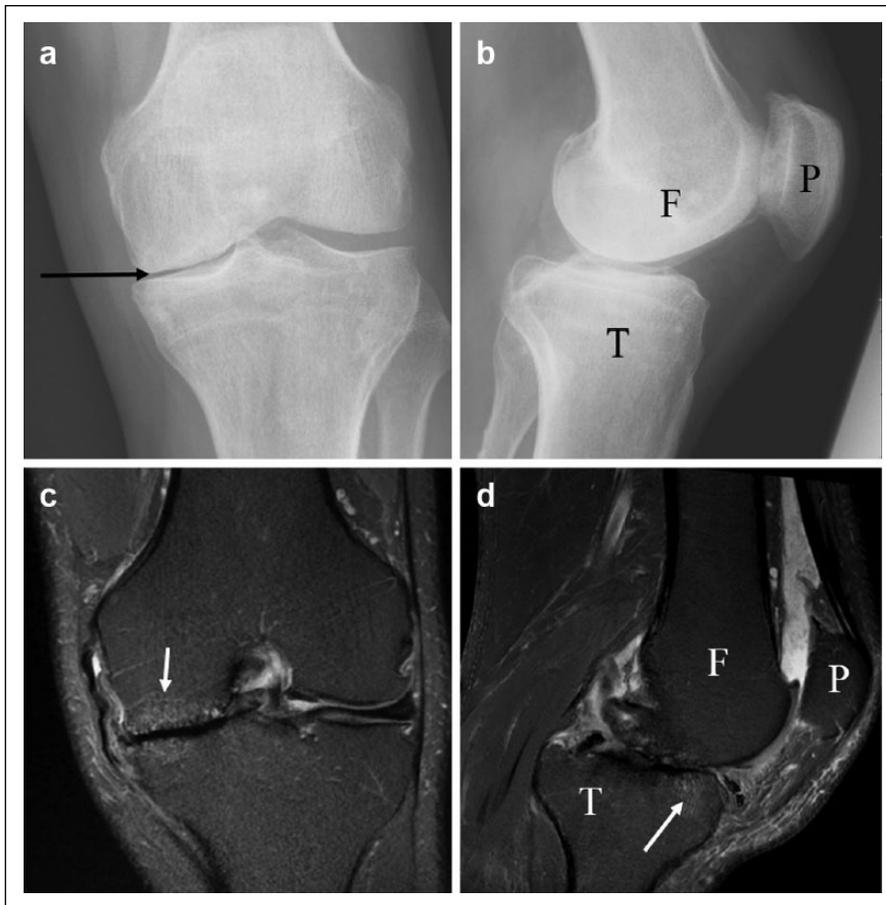


Figure 1. Radiographic and magnetic resonance (MR) images of the left osteoarthritic knee from a person with OA. The anterior-posterior (AP) projection radiograph (a) shows a narrowed medial tibiofemoral joint space (black arrow), whereas abnormalities are less apparent in the lateral view (b). In the MR images, bone marrow lesions can be seen in both (c) AP and (d) sagittal images (white arrows). F = femur; T = tibia; P = patella. Image credit: T. Kurien.

joint pain in univariate analyses, but independent associations with symptoms focus attention on cartilage integrity, synovitis, and subchondral bone as playing pivotal roles. Ultrasound-defined synovitis (**Fig. 2**) (effusion, synovial hypertrophy, and increased blood flow demonstrated by power Doppler signal) is also associated with OA pain, partly independent of radiographic change.¹³

Current Limitations of Existing Therapies

Major limitations of existing therapies include limited efficacy, benefit for only a proportion of patients, the need for continuous treatment to maintain benefit and because of the waning effectiveness with time from treatment initiation, and potential for important adverse events. Nonsteroidal anti-inflammatory drugs (NSAIDs) can be helpful, but importantly, they increase risks of ischemic cardiovascular events and can impair renal function. Increased risk of gastrointestinal bleeding with NSAIDs requires coadministration of gastroprotective agents. Paracetamol has limited benefit and might itself be associated with gastrointestinal blood loss.¹⁴ Opiates also have little sustained benefit for OA pain, and are associated with important gastrointestinal and cognitive adverse events, as well as their abuse potential. Local treatments, such as topically or

intra-articular drug administrations, might circumvent some of these limitations, but even topical NSAIDs or capsaicin have limited benefits, and intra-articular treatments have limited duration of benefit, raising logistic issues for repeated administration.

There are no medical disease-modifying treatments consistently proven to reduce structural OA changes, and the most effective structural modifying treatment is surgical arthroplasty, which itself is expensive and might only have an 80% success rate in relieving knee pain.¹⁵ Demonstration of disease-modifying activity in OA requires expensive and protracted clinical trials with large numbers of patients, due to the relatively slow and unpredictable rates of structural progression experienced by most people with OA, and the weak association between joint structure and clinically important outcomes, such as pain.

Limitations of preclinical models have also hampered the development of new medical treatments for OA pain. Animal models and behavioral tests used for developing OA analgesics are discussed in more detail elsewhere.¹⁶ Rodent models of OA pain are useful for drug development, although large animal models might better reflect some of the biomechanical influences in human OA. Pain behavioral techniques are well validated in rodent models, but more

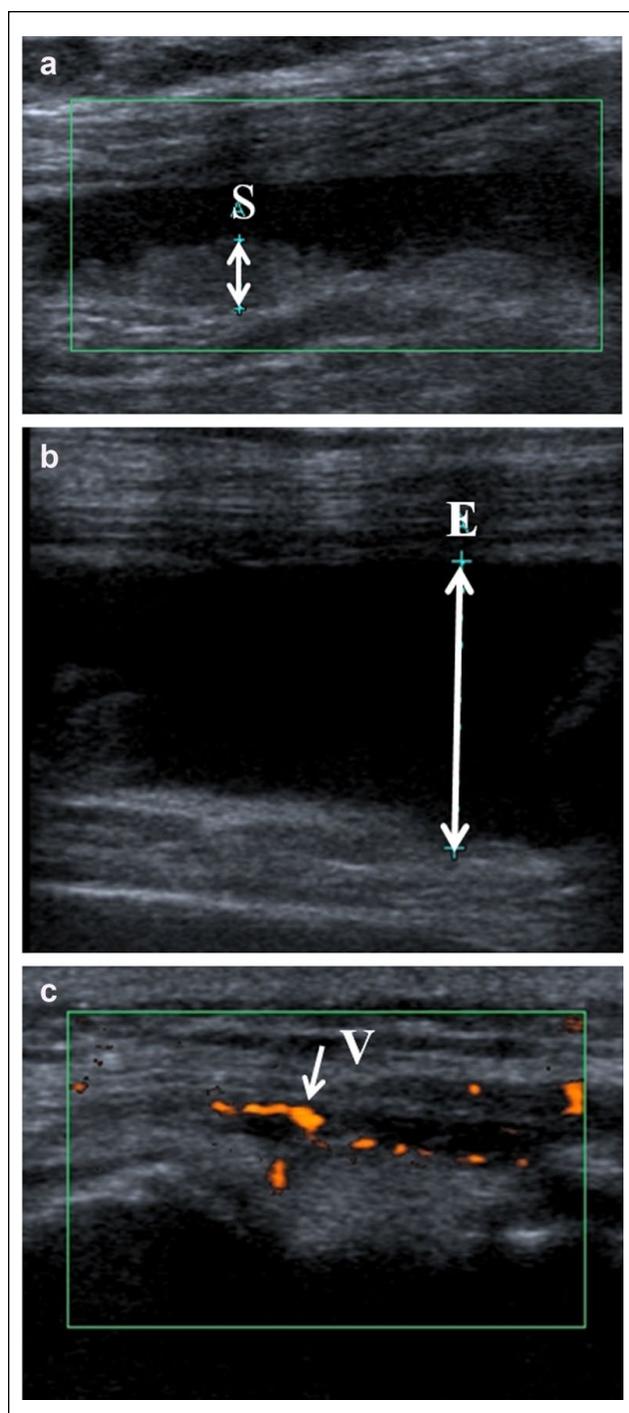


Figure 2. Ultrasound images of the knee of a person with OA: (a) synovial hypertrophy (S), (b) increased depth of synovial effusion (E), and (c) increased vascularity (V) demonstrated by power Doppler signal. Image credit: A. Sarmanova.

reproducible in rats than in mice, and more difficult in guinea pigs. OA is a disease of late maturity, and there remain concerns about the translational validity of induced OA in immature rodents. Surgical OA induction might better reflect initiating factors in at least some human OA (e.g.,

that following meniscal injury), whereas chemical induction, for example, by intra-articular injection of monoiodoacetate, provides a simpler method for rapidly developing a model of late-stage OA in rats. Different models might reflect different aspects of human OA, but there is no single model that reflects all aspects of human OA pain, and multiple models might need to be used to facilitate the generalizability of findings to human OA.

Modern Perspectives on OA Pain: The Quest for New Treatments

Current mechanistic understanding of the biochemical and cellular mechanisms of OA pain has led to optimism for the development of new and better treatments. More than 500 worldwide registered patents were filed between January 2014 and June 2016 related to OA therapies and biomarkers (Table 1). The focus of the majority of these patents can be categorized into treating inflammation, bone and cartilage, or sensory nerves. Novel tissue engineering and cell therapies continue to aim to protect structural integrity. Additional proposed treatments include traditional Chinese medicine or dietary supplements, although these often lack clear biomedical mechanistic rationale.

Inflammation and OA Pain

Synovitis in OA differs from that in RA, reflecting in OA predominantly innate rather than acquired immune responses, characterized by macrophage rather than lymphocyte infiltration.¹⁷ Correspondingly, the cytokine profile from an inflamed OA joint differs from that seen in RA, although some characteristics are shared between diagnoses, albeit usually at a lower intensity in OA. OA and RA synovial fluids each contain increased concentrations of tumor necrosis factor (TNF) α and interleukin (IL)-1, as well as growth factors, including nerve growth factor (NGF), transforming growth factor (TGF) β , and vascular endothelial growth factor (VEGF).¹⁸ Both display lipid-derived inflammatory mediators, such as E series prostaglandins.¹⁹ Active RA is characterized by a systemic acute phase response, with raised erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP). Acute phase response is also increased in people with OA compared with nonarthritic controls, although usually only to levels within accepted laboratory normal ranges.²⁰ Synovial fluid and serum cytokine concentrations, and serum acute phase response in OA, have each been associated with OA pain intensity (Table 2).²⁰ However, intercorrelation between inflammatory biomarkers prevents definitive elucidation of which specific markers might mediate OA pain.

Oral NSAIDs have long been known to reduce OA pain, and topical application might reduce pain from some arthritic joints, for example, hands or knees.²¹ Glucocorticoids by local injection have analgesic benefit in knee OA,²¹ but effects are temporary and repeated administration risks exacerbating

Table 1. Osteoarthritis and Biomarker Patents Registered between January 2014 and June 2016 Identified on <https://worldwide.espacenet.com> (June 2016) (further patent details can be found in the supplementary reference table).

| Inflammation | Patent Publication Number |
|--|---|
| Anti-IL1 | CA2931978 (A1), AU2014360704 (A1), MX2013009130 (A), MY155269 (A) |
| IL-1-RA adenoviral vector | INI682KON2014 (A), CN104245941 (A) |
| Autotaxin inhibitors | MX2015014298 (A) |
| NSAIDs | CN105198747 (A) |
| PGE receptor antagonists | IN719CHN2014 (A) |
| RGD proteins | INI981DEN2012 (A), CN104530233 (A) |
| GM-CSF | JP2015143233 (A), AU2015224416 (A1) |
| ICE inhibitor | US2015284426 (A1) |
| Anti- α 9 integrin | IN266863 (B) |
| Lubricin | US2015275118 (A1) |
| PEDF | TW201412327 (A), TWI491407 (B), WO2014043871 (A1), CN104903344 (A) |
| Anti-RGD antibodies | CN104530233 (A) |
| IL-17 inhibitors | NZ613656 (A), NZ613592 (A) |
| IL-18 inhibitors | WO2014023952 (A3), WO2014023952 (A2) |
| IFN- γ | NZ613656 (A), NZ613592 (A) |
| EGF receptor | CN104257659 (A) |
| PGE synthase-1 inhibitors | MX2014015350 (A), WO2015059618 (A1), JP2015172039 (A) |
| Anti-TIMP4 | WO2015116689 (A1) |
| Glucocorticoids | US2015141388 (A1) |
| Antifibrotic agents | WO2016044153 (A1) |
| TNF modulator | AU2015221459 (A1) |
| Nalidixic acid | GB2516138 (C), GB2516138 (A), GB2516138 (B) |
| Bone/Cartilage | |
| Calcitonin | CA2928654 (A1) |
| MMP | AU2015224468 (A1), JP2015079000 (A), US2014342983 (A1), US9260707 (B2), JP2015010081 (A), CN104906081 (A) |
| Aggrecanase | CN104940944 (A) |
| ADAMTS5 | US2015368188 (A1), CA2900116 (A1) |
| Cathepsin B | JP2015051985 (A) |
| Cathepsin D | US2015361037 (A1), US2015361053 (A1), CA2898077 (A1) |
| BMP | US2015306179 (A1), US2015190469 (A1) |
| Doxycycline | WO2015153864 (A3), WO2015153864 (A2) |
| FGF-18 | WO2015124739 (A1), WO2015124735 (A1), WO2015124731 (A1), WO2015097236 (A3), WO2015097236 (A2), WO2015097233 (A1), US2015218637 (A1), WO2015124739 (A1), WO2015097236 (A3), WO2015097236 (A2), WO2015097233 (A1) |
| Strontium | CN104788586 (A) |
| TGF β | US2015139909 (A1), WO2016019225 (A1), US2014342983 (A1), US9260707 (B2) |
| Chondroitin sulfate | MX2015009544 (A), CN104788586 (A), US2016074429 (A1), IN9832DEN2014 (A), US2015258134 (A1), AU2013265350 (A1) |
| Sensory Nerves | |
| Fms, Kit, Flt3, TrkA, TrkB, TrkC kinase inhibitors | TW201544506 (A), MY153951 (A), US9296726 (B2), US2015051196 (A1), US2015166547 (A1), US9260437 (B2) |
| TRPV1 antagonists | MX2014000344 (A) |
| Trk inhibitors | US2015274725 (A1) |
| CGRP antibodies | US2015259415 (A1) |
| FGF-9 | CN104162148 (A) |
| T-type calcium ion channel | JP2015143255 (A) |
| Anti-NGF | NZ608560 (A), US2015050286 (A1), US9212222 (B2), MY153781 (A), IL186403 (A), US2014017235 (A1), US2014220023 (A1), US9353176 (B2) |

(continued)

Table I. (continued)

| Inflammation | | Patent Publication Number |
|-----------------------------------|---|---|
| | AMPA/KA glutamate receptor antagonists | EP3016636 (A1) |
| | H3 receptor antagonist | US2016089378 (A1) |
| | Capsaicinoids | PT1605956 (E) |
| | Botulinum toxin | KR20160002207 (A), US2016074485 (A1) |
| | Clostridial derivative | US2015086531 (A1) |
| Dietary supplements | Fish oil/cartilage | US2015306132 (A1), US2015231089 (A1), SG1120150021IP (A), EP2902033 (A1), US2014011888 (A1) |
| | Niacin | US9193708 (B2), US2015099787 (A1) |
| | Ginger | JP2015198661 (A) |
| | Curcumin/Vit K2 combinations | MX2015002013 (A) |
| | Tocotrienol | KR20150130749 (A) |
| Traditional Chinese medicines | | CN105267693 (A), CN105250392 (A), CN104383301 (A), CN104398803 (A), CN105343670 (A), CN105327144 (A), CN105233041 (A), CN105343700 (A), CN105012614 (A), CN104740120 (A), CN104225449 (A), CN104173511 (A), CN104225112 (A), CN104173667 (A), CN104225100 (A), CN104398710 (A), CN104523947 (A), CN105326928 (A), CN104524229 (A), CN104689176 (A), CN104306606 (A), CN104873673 (A), CN104352633 (A), CN105012531 (A), CN104474110 (A), CN104474110 (A), CN104547091 (A), CN104288673 (A), CN104721462 (A), CN105079721 (A), CN104667093 (A), CN104435314 (A), CN104324096 (A), CN104586942 (A), CN104324108 (A), CN105168711 (A), CN105192788 (A), CN104587059 (A), CN105079253 (A), CN104800823 (A), CN104547088 (A), CN105311490 (A), CN104324097 (A), CN104127856 (A), CN104127455 (A), CN104116976 (A), CN103520282 (A), CN103520282 (B), CN104606409 (A), CN104800267 (A), CN105169054 (A), CN105106466 (A), JP2015007037 (A), CN105012594 (A), CN104983920 (A), CN104922322 (A), CN104887965 (A), CN104666465 (A), CN104958520 (A), CN104606383 (A), CN104288410 (A), CN105148179 (A), CN104306647 (A), CN104666490 (A), CN105250392 (A), CN104510792 (A), CN104606411 (A), CN104587273 (A), CN104721366 (A), CN104784270 (A), CN104353064 (A), CN104983909 (A), CN104983908 (A), CN104983907 (A), CN105031029 (A), CN104645164 (A), CN104740568 (A), CN104815097 (A), CN104906325 (A), CN104352741 (A), CN104887869 (A), CN104706759 (A), CN104800673 (A), CN104523894 (A), CN105288016 (A), CN104873631 (A), CN104324322 (A), CN104984310 (A), CN104138455 (A), CN103536777 (B), CN103536777 (A), CN104116915 (A), CN103520654 (A), CN103520654 (B), CN104606585 (A), CN104873818 (A), CN104208587 (A), CN105194006 (A), CN103536755 (B), CN103536755 (A), CN104645056 (A), CN104306756 (A), CN104940319 (A), CN105106339 (A), CN104721644 (A), CN104540513 (A), CN104800802 (A), CN104548014 (A), CN104983825 (A), IN428CHN2014 (A), CN104922183 (A), CN104491485 (A), CN105147749 (A) |
| Tissue engineering/cell therapies | | WO2015124735 (A1), AU2014211790 (A1), IN2045MU2013 (A), CN104707140 (A), CN104706675 (A), CN105106238 (A), US2016000830 (A1) |
| Combinations | Hyaluronic acid/piroxicam composition | US2016106774 (A1) |
| | Corticosteroid/zoledronate coadministration | KR20150125001 (A), US2015031649 (A1), CN105324113 (A) |
| Miscellaneous | LNC RNA HI9 | CN105194690 (A) |
| | TMCO3 | CN105194674 (A) |
| | Anti-Fas antibody | CN105169389 (A) |
| | IK1 channel activator | TW201536775 (A) |

(continued)

Table 1. (continued)

| Inflammation | Patent Publication Number |
|---------------------------|---|
| Vit A/RARB agonist | US2015352070 (A1) |
| Coumarin derivative | US9365533 (B2), US2015232440 (A1), JP2015193610 (A) |
| Amylin peptides | TN2013000491 (A1) |
| Platelet-rich plasma | AU2014245854 (A1), WO2015003623 (A1), CN104707141 (A), US2016051701 (A1), CN104958320 (A), WO2015085957 (A1) |
| Myostatin | MX2015002894 (A) |
| CRAC channel modulators | NZ609572 (A) |
| CDK9 inhibitors | US2015105423 (A1) |
| Anemonin | CN104337804 (A) |
| CCDC59 agonist | CN105219877 (A) |
| Arginine/citrulline | EP3010496 (A1) |
| Indazole/indole/imidazole | IN2646CHN2014 (A), NZ610699 (A), CN105120862 (A), US9199991 (B2), US2015152105 (A1), CN105037355 (A), TN2013000434 (A1) |
| microRNA 101 | CN104083761 (A) |
| Zinc-ZIP8-MTF1 | US2015323528 (A1) |
| TSPAN 15 | CN105349644 (A) |

ADAMTSS, a disintegrin and metalloproteinase with thrombospondin motifs 5; AMPA/KA, Ampa kainate subtype of the glutamate receptor; BMP, bone morphogenetic protein; CCDC, coiled-coil domain containing; CDK, cyclin-dependent kinase; CGRP, calcitonin gene-related peptide; CRAC, calcium-release activated calcium; EGF, epidermal growth factor; Fas, tumor necrosis factor receptor superfamily, member 6; FGF, fibroblast growth factor; Flt3, Fms-related Tyrosine Kinase 3; Fms, feline McDonough sarcoma; GM-CSF, granulocyte-macrophage colony-stimulating factor; ICE, interleukin-1 converting enzyme; IFN, interferon; IK 1, inward rectifier current; IL, interleukin; IL-1-RA= interleukin 1 receptor antagonist; Kit, stem cell factor receptor; LNC RNA, long noncoding RNA; MMP, matrix metalloproteinases; MTF1, metalregulatory transcription factor-1; NGF, nerve growth factor; NSAID, nonsteroidal anti-inflammatory; PEDF, pigment epithelium-derived factor; PGE, prostaglandin E; RARB, retinoic acid receptor beta; RGD, Arg-Gly-Asp; TIMP, tissue inhibitor of metalloproteinases; TMCO3, transmembrane and coiled-coil domains 3; TNF, tumor necrosis factor; Trk, tropomyosin receptor kinase; TSPAN, tetraspanin; TPVI, transient receptor potential cation channel subfamily V member 1.

Table 2. Biomarkers of OA Pain, 2014–2016.

| Blood | Synovial Fluid |
|---|-----------------------------------|
| Increased | Increased |
| 2-AG ¹³⁹ | Autotaxin ¹⁴⁰ |
| Autotaxin ¹⁴⁰ | CCL2 ¹⁴¹ |
| BDNF ¹⁴² | CD14 ¹⁴³ |
| CB1R/CB2R (PBL expression) ¹³⁹ | CGRP ¹⁴⁴ |
| CD14 ¹⁴³ | CX3CL1/fractalkine ¹⁴⁵ |
| CGRP ¹⁴⁴ | YKL-40 ^{146,147} |
| COMP ¹⁴⁸ | |
| CX3CL1/fractalkine ¹⁴⁵ | Decreased |
| HMGB-1 ¹⁴⁹ | IL-17 ¹⁵⁰ |
| sHA ¹⁴⁸ | sTNFR1, sTNFR2 ¹³⁶ |

2-AG, 2-arachidonoylglycerol; BDNF, brain-derived neurotrophic factor; CB1R/CB2R, cannabinoid receptors 1 and 2; PBL, peripheral blood leukocyte; CCL, C-C motif chemokine ligand; CD14, cluster of differentiation 14; CGRP, calcitonin gene-related peptide; COMP, cartilage oligomeric matrix protein; CX3CL1, fractalkine; HMGB, high-mobility group box; IL, interleukin sHA, serum hyaluronic acid; sTNFR, soluble tumor necrosis factor receptor; YKL-40, chitinase-3-like protein 1 (CHI3LI).

structural damage, for example, through avascular necrosis. Systemic corticosteroids have not shown consistent benefit in OA,²² and are largely precluded by adverse events with long-term use. Other drugs known to have anti-inflammatory

disease-modifying activity in RA (hydroxychloroquine or methotrexate) offer some promise in OA, and the results of ongoing randomized controlled trials (RCTs) are eagerly awaited.^{23–26} TNF α is produced by synovium both in RA and in OA, and antibodies that block TNF α activity substantially reduce pain from active RA.²⁷ However, benefits have been inconsistent from TNF α blockade in OA,^{28–30} whether administered systemically or by intra-articular injection, and other inflammatory mediators might predominate in OA.

Recent research has further explored the potential of inflammation-related biomarkers to predict OA pain, including soluble CD14 and hyaluronic acid (Table 2). Furthermore, inflammatory mediators are subject to intensive investigation as possible therapeutic targets that might both reduce pain and reduce inflammatory drive to joint damage (Table 1). Recently patented agents have targeted IL-1, -17, or -18, and TNF α , as well as trying to improve targeting of more traditional OA treatment targets, such as prostaglandins and corticosteroid pathways.

Osteochondral Change and OA Pain

Subchondral bone is attracting increasing attention as a potential treatment target for OA pain. MRI-defined BMLs are associated with pain in cross-sectional and prospective studies. BMLs represent regions of increased metabolic

activity and increased bone turnover,³¹ and correspondingly, OA is associated with increased osteoclast activity within subchondral bone, both in man³² and in animal models.³³ A range of osteoclast inhibitors have demonstrated analgesic efficacy in preclinical studies, including bisphosphonates,³⁴ osteoprotegerin,³³ and cathepsin K inhibitors.³⁵ Preliminary evidence from randomized clinical trials, which often focused on possible structural disease-modifying activity, suggest analgesic efficacy.^{36–38} Efficacy in unstratified cases with large joint OA was inconsistent, but based on the assumption that osteoclast inhibition should only be effective in the subgroup of people with OA who had increased bone turnover, Laslett et al. reported an RCT of zoledronate in people with knee OA and current BMLs on MRI scans at recruitment.³⁶ A single infusion of zoledronate was followed by significant pain reduction at the primary end point of 6 months. Another RCT, using neridronate, further suggests similar analgesic activity,³⁷ and a definitive trial of zoledronate is due to be completed in 2017. Strontium, which has mixed anabolic and anticatabolic activity on bone, also reduced BMLs³⁹ and showed pain-reducing effects in human OA,⁴⁰ although potential cardiovascular events might limit implementation of these findings into clinical practice.

Recent patents aiming to develop novel bone-targeted treatments might also realize the potential to reduce symptoms, while avoiding the need for prolonged and expensive clinical trials aiming to modify structural progression (**Table 1**). Many of these agents, such as strontium, might also have structural effects mediated through chondrocyte function.

Mechanotransduction

OA pain is characteristically both intermittent and constant.⁴¹ Intermittent and constant pain reflect discrete domains, as determined by Rasch analysis of questionnaire data,⁴² and therefore are likely to be underpinned by discrete mechanisms. Intermittent OA pain is characteristically mechanically transduced, for example, on weight bearing or during walking, and mediated by fast-conducting myelinated nerve fibers. Precise molecular pathways of nociceptive mechanotransduction are incompletely understood, but likely involve ion channels, such as TRPA1,⁴³ short transient receptor potential channel 3 (TRPC3),⁴⁴ piezo 2,⁴⁵ and others.⁴⁶ Mechanotransduction also requires interaction between nerves and ancillary cells.⁴⁶ Inhibiting mechanotransduction as a means of reducing OA pain is an attractive proposition, commonly exploited by nonpharmacological interventions that redistribute mechanical stresses across the joint, for example, through orthoses⁴⁷ or by surgical osteotomy.⁴⁸ Pharmacological manipulation of mechanotransduction has presented more challenges to date. Mechanotransduction is also important for hearing and blood pressure control, and genetic deletion of TRPC3⁴⁴ or acid-sensing ion channel 2 (ASIC2)⁴⁹ mechanotransducing

molecules results in hearing-impaired or hypertensive phenotypes, respectively. Identifying molecules with a specificity for nociceptive mechanotransduction, however, remains a desirable objective for pharmaceutical development.

Peripheral Nervous System

Nociceptive transmission, in either peripheral or central nerves, offers another potential target for treating OA pain. Intra-articular lignocaine injection can produce temporary analgesia,⁵⁰ presumably in part through nonselective sodium channel blockade (although also through placebo effects⁵¹). Sodium channel subtypes involved in nociceptive pain include Nav1.7, and antagonists of Nav1.7 have displayed analgesic efficacy in preclinical models.⁵² Clinical development of Nav1.7 inhibitors has, however, been limited by adverse events, including arrhythmia, seizures, and sedation, probably in part through cross-reactivity with other sodium channel subtypes (e.g., Nav1.6). Greater selectivity for Nav1.7, and peripherally restricted antagonists that less easily cross the blood–brain barrier are entering clinical development for neuropathic pain.

The clinical relevance of peripheral sensitization to OA pain has been confirmed by the analgesic efficacy of three different NGF-blocking antibodies (tanezumab, fulranumab, and fasinumab) in people with OA,⁵³ and additional NGF-blocking antibodies are poised to enter clinical development.⁵⁴ NGF pathways and neurotrophin receptors, such as tropomyosin receptor kinase (Trk) A, represent a high proportion of recent patents for novel OA treatments (**Table 1**). Analgesia with NGF-blocking antibodies has been substantial (effects above placebo greater than observed with NSAIDs) and sustained during repeated administration over months. Efficacy has been demonstrated in knee⁵⁴ and hip OA⁵³ and also in low back pain,⁵⁵ a condition notoriously resistant to analgesic medications in RCTs and clinical practice. Other approaches to NGF pathway inhibition have focused on small molecular inhibitors of TrkA kinase. Development has been frustrated by lack of specificity for TrkA over other kinase receptors, but recent small-molecule allosteric TrkA inhibitors have been described with >100-fold selectivity for TrkA over TrkB and TrkC, and even greater specificity over other tyrosine kinase receptors. Preclinical data suggest that nonselective or selective TrkA inhibition can reproduce the analgesic effects of NGF-blocking antibodies⁵⁶ (compare efficacy with data in Xu et al.⁵⁷).

The precise molecular and cellular mechanisms by which peripheral sensitization contributes to OA pain remain unclear, raising the possibility that specific molecular targets other than TrkA might also generate effective analgesic agents. TrkA is coexpressed with transient receptor potential cation channel subfamily V member 1 (TRPV1) and calcitonin gene-related peptide (CGRP), on sensory nerves that also express CGRP

receptors. NGF induces phosphorylation, increased expression, and thereby increased activity of TRPV1.⁵⁸ Small-molecule TRPV1 inhibitors did not display OA analgesic activity in RCTs, although adverse reactions, both on target (e.g., hyperthermia) and possibly off target (e.g., hepatotoxicity), might have precluded the use of doses adequate for clinically important pain relief. Further, TRPV1 inhibitors that are selective for sensory nerve function rather than thermoregulation continue in development. NGF also increases CGRP expression, which itself contributes to peripheral sensitization in rat OA models.⁵⁹ Small-molecule CGRP receptor antagonists were similarly limited by liver toxicity, but more recently, CGRP-blocking antibodies have demonstrated efficacy in RCTs as prophylactic agents against migraine,⁶⁰ although an RCT of one CGRP-blocking antibody in people with OA pain was disappointingly negative.

Central Nervous System

People with OA frequently display evidence of central sensitization: augmented central nociceptor activity in response to normal primary afferent inputs. Increased temporal summation (augmented pain intensity felt on repeated application of an equal-intensity stimulus), increased intensity and distribution of secondary hyperalgesia around a focus of primary insult, and augmented receptive fields and amplitude of flexor withdrawal reflexes each suggest spinal sensitization by analogy with similar phenomena observed in experimental animals.⁶¹ Spinal sensitization is partly mediated by *N*-methyl-D-aspartic acid (NMDA) receptor activation,⁶² but pharmacological NMDA receptor blockade, for example, with ketamine, is associated with adverse events that have precluded clinical application in OA.

OA pain phenotypes are in part dependent on the integrity of central inhibitory and facilitatory pathways originating above the spinal cord. People with OA frequently display impaired conditioned pain modulation (CPM), whereby a second painful stimulus fails to suppress pain intensity elicited by a test stimulus. Blunting of CPM might reflect impaired descending inhibition and/or increased descending facilitation. Descending control is, in part, mediated by serotonin⁶³ and dopaminergic⁶⁴ pathways, and serotonin/noradrenaline reuptake inhibitors are most likely to display analgesic benefit for neuropathic pain in those with impaired CPM.⁶⁵ Pregabalin, despite demonstrating benefit in other centrally mediated pain states, such as fibromyalgia,⁶⁶ displayed an absence of analgesic benefit in one RCT of people with hip or knee OA.⁶⁷ This might indicate that central pain augmentation is a major contributor to OA pain in only a minority of patients. Duloxetine, a selective serotonin/nor-epinephrine reuptake inhibitor (SSNRI), displayed significant analgesic activity in people with OA,⁶⁸ although overall effect sizes were small compared with placebo, again possibly reflecting response heterogeneity between individuals. Placebo analgesia might also offer the potential to activate endogenous analgesic pathways through psychological (e.g.,

cognitive) mechanisms. Indeed, placebo analgesic effect sizes exceed pharmacological effects in RCTs of most analgesic agents tested in OA.⁶⁹ If duloxetine and pregabalin share analgesic mechanisms with placebo, comparisons against placebos in RCTs might underestimate their full clinical potential.

OA pain is both a sensory and emotional experience, and negative affect, reflecting an overlapping construct of anxiety and depression, is strongly associated with musculoskeletal pain.⁷⁰ Indeed, brain functional and connectivity signatures of OA pain are localized to brain structures, such as the periaqueductal grey and anterior cingulate cortex,⁷¹ regions known to be involved in mood disturbance.^{72,73} Negative affect is associated with impaired CPM^{74–76} and reduced (more sensitive) pressure pain detection thresholds,⁷⁷ in people with or without OA.

Modulation of anxiety or low mood, either pharmacologically or through cognitive behavioral therapy, has the potential to improve not only the emotional component of OA pain, but also the alterations in central pain modulation that can augment its sensory component.^{65,78} However, not all antidepressant or anxiolytic classes have demonstrated analgesic efficacy in OA. Early research suggested that depression in people without chronic pain might be associated with higher (less sensitive) pain thresholds,⁷⁹ but interactions between negative affect and chronic pain and a primary drive to pain sensitization from anxiety or depression remain to be fully elucidated. Strong associations between catastrophizing,⁸⁰ or fear avoidance beliefs,⁸¹ and current or future joint pain⁸⁰ might suggest a primary association with anxiety rather than depression.

Ongoing nociceptive barrage is believed to facilitate and maintain central sensitization, although systemic or chemical factors might also contribute.⁸² Several analgesic strategies have been found to reduce evidence of central sensitization, most notably joint replacement surgery. Widespread reductions in pressure pain detection thresholds⁸³ and impaired CPM,^{83,84} prominent in people with knee or hip OA, can each normalize after successful joint replacement surgery. However, associations between these clinical correlates of central sensitization and pain are, in part, genetically determined,^{63,85,86} and might therefore precede OA onset, indicating that nociceptive input is only one of several factors leading to central pain augmentation. Furthermore, an important minority of people continue to experience pain after arthroplasty (estimated at up to 20% after knee arthroplasty and 5% after hip arthroplasty) and continue to display evidence of central sensitization,⁸⁷ suggesting that once developed, central sensitization might become independent of ongoing nociception.

A direct contribution of synovitis to the development of central sensitization has been suggested by clinical and pre-clinical studies. MRI evidence of synovitis, but not BMLs, was associated with reduced pressure pain detection thresholds in one cross-sectional study of people with OA.⁸⁸

Widespread reductions in pressure pain detection thresholds have been associated with serum acute phase response in human OA.^{89,90} In rodent models of OA pain, synovitis is followed by evidence of central sensitization.⁹¹ It remains possible that cytokines or growth factors, other than those classically recognized as driving inflammation, might generate central sensitization. The neurotrophin NGF increases brain-derived neurotrophic factor (BDNF) expression and central release by dorsal root ganglion (DRG) cells, contributing to central sensitization.⁹² The potential of anticytokine treatments or neurotrophin blockade to inhibit the development or maintenance of central sensitization, particularly in patients with inflammatory OA, remains to be fully explored.

Glial cell activation in the spinal cord (first microglia, later accompanied by astrocytosis), and subsequently in the brain, develops in parallel to central sensitization in animal models of OA, and administration of glial cell inhibitors reduces sensitization.⁹³ Positron emission tomography scans using radiolabeled antagonist ligand to translocator protein (TSPO) suggest increased glial activity in brains of people with chronic low back pain,⁹⁴ as do increased cerebrospinal fluid (CSF) concentrations in OA or other musculoskeletal pain of molecules (e.g., ILs and cystatin C) known to be expressed by glial cells.^{95,96} Some existing analgesics might inhibit glial cell activity,⁹³ and novel, centrally acting analgesics that might reduce glial-mediated central sensitization have considerable potential for OA pain.

OA-specific drug patents lag behind the recent increases in understanding of central pain mechanisms in OA (**Table 1**). These central pain mechanisms might not be OA specific, but rather shared, for example, with neuropathic pain. Analgesics under development for nonmusculoskeletal conditions might therefore help at least some people with OA in the future.

Systemic Influences on OA Pain

Systemic factors also contribute to OA pain. Structural and painful OA are associated with body mass index (BMI), and weight reduction is associated with reduced pain, although the effect size of dietary intervention is small.⁹⁷ Associations between BMI and OA pain might partly be explained by increased biomechanical stress, although associations between BMI and OA hand pain,⁹⁸ and indeed between BMI and non-OA pain (e.g., fibromyalgia⁹⁹), point to additional pain mechanisms. BMI and OA pain might be linked genetically, and adipocytokines circulating at higher levels in obese patients might contribute to neuronal sensitization.^{100,101} Identifying the mechanisms that link BMI to OA pain could lead to more refined and effective analgesic treatment than can realistically be achieved by dieting alone.

Treatment Strategies

Two areas of intensive research are the use of combination therapies and the development of robust biomarkers to help design and target effective OA pain treatment. Both have the potential to increase benefit from existing therapies and go hand in hand with the development of novel treatments.

Combination Therapies

Given the multiple contributing mechanisms in OA pain, it is unsurprising that no single treatment has proved efficacious for all patients. Combination therapy is the norm. Pharmacological interventions are typically delivered within the context of other treatment modalities, including psychological and physiotherapeutic approaches. Combination pharmacotherapy is commonly used, despite little robust evidence of greater benefit over monotherapies, and despite potential for drug interactions and adverse events.

Combination pharmacotherapy might most likely be effective where drugs act through discrete mechanisms. Combining two NSAIDs should have no greater than an additive benefit, and combining full and partial agonists might be expected to *reduce* benefit. Regular use of sustained-release or long-acting analgesics is recommended for chronic, constant pain. However, patients often choose to use analgesics intermittently, or use top-up analgesia to treat pain flares. People use analgesics prophylactically to facilitate activities that are anticipated to be painful. Caution is indicated, however, in ensuring that treatment combinations genuinely meet patients' needs rather than concealing abuse of prescribed medications. Combination therapies might also incorporate nonpharmacological interventions. For example, effective analgesia might facilitate engagement in exercise, and analgesic drugs might therefore synergize with physiotherapy. All medications should be provided together with matched cognitive reassurance.¹⁰²

RCTs of combination therapy are difficult, expensive, and rare. An ideal trial design might require several parallel intervention arms in a double-dummy design: double placebo versus monotherapy A + placebo versus monotherapy B + placebo versus combination therapy A + B. Clinical practice tends to adopt a step-up approach, in which case the sequence of interventions requires randomization, further increasing trial complexity. RCTs testing combination pairs of acetaminophen (paracetamol), NSAIDs, and/or opioids have been reported, although clinically important benefits of combination over monotherapy with one or both components have not been consistently demonstrated.¹⁰³ Recent evidence that paracetamol might share cyclooxygenase inhibition with NSAIDs, as well as risk of gastrointestinal blood loss,¹⁰⁴ raises concerns about paracetamol + NSAID combinations.

Unexpected adverse events might result from pharmaceutical combinations. Phase III RCTs of NGF-blocking antibodies revealed rapidly progressive OA (RPOA) at increased

frequency in people taking NSAIDs concurrently with NGF-blocking antibodies.¹⁰⁵ NSAIDs alone have been associated with RPOA, and retrospective analysis of trial data indicated a synergistic and dose-dependent effect of added NGF blockade.¹⁰⁶ The mechanism of this interaction remains uncertain, and both actions on bone turnover⁵⁷ and joint usage¹⁰⁷ in the context of analgesia have been proposed.

Monotherapies acting through discrete molecular mechanisms on central pain processing have demonstrated small clinical benefits in RCTs in OA, but it is unknown whether their combination with peripherally acting drugs would be more effective than either treatment alone. Combinations of duloxetine plus gabapentinoid in diabetic neuropathy have shown marginal possible benefits compared with either alone,¹⁰⁸ but similar combinations have not been reported in OA. Epidemiological evidence has suggested that concurrent treatment with beta-blockers, such as propranolol, is associated with lower reported pain and lower opiate use in people with OA.¹⁰⁹

Monotherapies might be tried before combinations, and treatments should only be continued if providing a benefit, and continued at the lowest effective dose in order to minimize risk of adverse events. Possible synergy between analgesic drugs, however, means that failure to detect a clinically important benefit from monotherapy might not exclude potential benefit in combination. A “real-life” trial protocol in which patients only progress to combination therapy if they display partial response to monotherapy might conceal synergistic benefit from combinations. Furthermore, failure to respond to one analgesic agent might predict failure to respond to a subsequent analgesic, most obviously where both act through the same mechanism. The converse observation, in that those who respond to one treatment might be more likely to respond to another in the same class, has informed flare RCT designs where participants are enriched with responders by identifying those whose pain flares following withdrawal of their regular analgesic.¹¹⁰ However, psychological factors also contribute to therapeutic failure. When expectations of pain are incongruent with repeated experience, alterations in central pain processing can either augment or suppress pain.¹¹¹ Repeated exposure to ineffective treatments (especially where associated with adverse events) might create an expectancy of nonresponse, thereby blunting placebo and augmenting nocebo effects. A step-up approach might therefore lead to underestimation of benefit from combination therapies, and should ideally be tested in RCTs against concurrent or step-down approaches.

Biomarkers and Treatment Targeting for OA Pain

Biomarkers have potential utility to help diagnose patient subgroups, measure disease burden, define prognosis, and predict treatment outcomes (efficacy and adverse events).¹¹² Patients with highest disease burden, or whose disease is likely to get worse, stand to gain most from treatment, and

are therefore likely to display the highest likelihood of benefit over risk. Several biomarkers have provided evidence of identifying those most likely to experience progressively painful OA, including collagen degradation products, urinary cross-linked C-telopeptide (uCTX) v II and uCTXI α ,¹¹³ and serum cartilage oligomeric matrix protein (COMP).¹¹⁴ Overall however, the predictive value of existing biomarkers, either alone or in combination, is weak. Evidence that a biomarker might predict prognosis might currently be more useful in pointing to possible disease mechanism rather than indicating tools for stratifying treatments.

Given the complexity of pain pathways, their numerous modulating factors, and the clinical heterogeneity of OA, it seems likely that different treatments might be effective for different people, for different aspects of their pain problem or at different stages of their disease. Stratification of patients to select groups that are most likely to benefit from treatment has potential to improve success in clinical trials of analgesics with specific modes of action. Recruiting from homogenous groups of people who are most likely to respond to treatment permits increased statistical power, reduced trial size, and reduced exposure of those unlikely to respond to potential adverse effects of treatment. Offering to patients the treatments to which they are most likely to respond, at the earliest point in their care pathway, should reduce the suffering and disengagement inherent in cycles of ineffective treatment. However, individuals with OA might represent points on a continuum, each with a unique probability of response but without definable boundaries that would justify treatments being offered to (or withheld from) discrete patient groups.

Predicting treatment response from pretreatment characteristics is further hampered by the observation that only approximately 25% of overall treatment effect is attributable to specific pharmacological activities rather than placebo response.¹¹⁵ Who will or will not benefit from a treatment might therefore be better predicted by determinants of placebo response rather than by matching the pharmacological target to an individual's pain mechanism. Predictors of placebo response differ substantially from factors that predict pharmacological response, and include genetic, gender, and personality characteristics. Accumulating risk of pharmacological adverse events with long-term treatment of OA pain might not be justified if analgesic mechanisms are due to treatment context, particularly if placebo analgesia might be achieved by other methods without pharmacological risk. Pharmacological benefit to an individual should ideally be determined both in clinical trials and in clinical practice in order to justify progression to long-term treatment of chronic arthritis pain. Distinguishing pharmacological from placebo response, however, would require robust and validated biomarkers of treatment effects, or multiple crossover, double-blind, placebo-controlled trial designs. Such methodologies have generally not been translated into clinical practice, and rarely inform individual decision making on treatment continuation.

Clinical history and examination provides the cornerstone of patient stratification but has limited sensitivity or specificity for predicting treatment response for OA pain. Those with the most severe symptoms stand to gain most from treatment, and treatments with the highest risk or cost are generally reserved for those with the worst disease. However, where clinical risk factors for treatment outcome have been identified, their predictive value might not be sufficient to stratify treatment allocation. For example, high BMI predicts worse outcome from arthroplasty, but arthroplasty remains a cost-effective treatment option for those with high BMI.¹⁴ Clinical examination displays only weak correlation with imaging evidence of synovitis, and poorly predicts response to anti-inflammatory treatments, such as intra-articular steroid injections.¹¹⁶ Biomarkers that are more robust are required to inform clinical decision making.

Genetic Biomarkers

Genetic differences between individuals explain up to 40% of OA pain,¹¹⁷ representing interactions between a large number of genetic loci, each alone contributing little predictive value. Targeted gene approaches have provided evidence for contributions from specific genes in general OA populations, and genome-wide association studies suggest additional, as yet uncharacterized, loci. Genetic risk factors for OA pain include risk of OA structural change, including bone and cartilage matrix integrity and shape, and inherited variability in pain processing. Inheritance segregates between hip, knee, or hand OA, highlighting constitutional risk factors for OA subgroups. Specific associations with allelic variation in *Nav1.7* and TRPV1 ion channels, P_2X_7 purinergic receptors, catecholamine *O*-methyl transferase (COMT), and paired amino acid converting enzyme 4 (PACE4)^{117,118} might be shared with other chronic pain conditions, including neuropathic pain, and reflect heterogeneity in pain neurotransmission. Genetic variation might underpin differences in emotional¹¹⁹ or descending pain modulation,⁶³ implicating, for example, serotonin transporter polymorphisms. Genetic variants also predict placebo responses,¹²⁰ analgesic (e.g., opioid) responses, or adverse events.¹²¹ The use of genotypes (either individually or in combinations) to select people with favorable risk–benefit ratios for specific treatments remains in its infancy in OA. Examples in other branches of medicine have tended to be most successful with variants in single genes, as exemplified by the thiopurine methyltransferase genotype predicting toxicity risk from azathioprine. The polygenic nature of OA pain makes genotypic stratification more challenging.

Wet Biomarkers

Markers measured in biofluids have offered additional hope in subgrouping people with OA by pain mechanisms (Table 2). Markers of synovitis have shown some promise, although systemic inflammatory responses are much less in

OA than in RA. Any single, symptomatic joint might contribute little to circulating biomarker levels. Synovial fluid cytokine levels have been more often associated with OA pain than blood biomarkers, but synovial fluid volumes might be small, and aspiration might not be a practicable procedure for routine use in the clinical management of OA. ESR and serum CRP might each be increased in OA compared with nonarthritic controls, although usually still within the “normal” range.²⁰ Associations between serum’s acute phase response and OA pain might reflect a contribution from synovitis, but might otherwise be explained by known associations between CRP and high BMI.¹²² Reduced leptin and IL-6 were associated with decreasing OA pain, in parallel to weight reduction following a behavioral weight management¹²³ intervention. Serum levels of various cytokines have also been associated with OA pain (Table 2), mostly in cross-sectional studies. Increases in serum cytokine levels over time have less frequently been associated with increasing OA pain.

Serum or urine biomarkers of cartilage turnover, including COMP (Table 2), have predicted structural and pain progression in OA.^{113,114} Serum biomarkers of osteoclast activity have been developed for partnering osteoporosis treatment. OA pain has been associated with increased urinary degradation products of collagen I,¹¹³ the predominant bone collagen, although also a constituent of other connective tissues. Tartrate-resistant acid phosphatase type b5 (TRAP5b), released by osteoclasts during bone degradation, can be measured in serum by enzyme-linked immunosorbent assay. Serum TRAP5b was associated with subchondral osteoclast numbers in OA knees at arthroplasty, and, in a nonsurgical cohort, was associated with current OA pain and predictive of pain at follow-up.¹²⁴ Circulating bone and cartilage biomarkers, however, have displayed only weak correlations with current symptoms, perhaps reflecting origins from tissues in addition to those joints that are affected by symptomatic OA.

Imaging Biomarkers

Imaging biomarkers have thrown light on OA pain subgroups. Joint space narrowing on posteroanterior weight-bearing radiographs are associated with OA pain.¹²⁴ MRI scans also indicate associations between pain and cartilage thickness or surface area, and also with meniscal morphology.^{125–128} Each of these MRI features is associated with joint space narrowing on plain radiographs. Arthroplasty provides a mechanical and chemical barrier between joint and subchondral bone, and might be most effective when that barrier has been disrupted by late-stage OA. Indeed, people with less preserved radiographic tibiofemoral joint space are more likely to benefit from total joint replacement surgery.¹³⁰ Other mechanisms might drive pain in people with OA in whom joint space is preserved, mechanisms that might persist after arthroplasty. Up to 20% of patients continue to experience disabling knee pain after arthroplasty, and ongoing research is determining the

extent to which this is due to pain mechanisms that persist despite surgical treatment (e.g., augmented central pain processing⁸⁷), or to sequelae of joint surgery (e.g., neuropathic pain from intraoperative nerve damage¹³¹).

Imaging evidence of synovitis predicts response to intra-articular glucocorticoid treatment. The predictive value of imaging is weak, although better than evidence of synovitis from clinical examination. Reductions in synovitis, for example, determined by a reduced early enhancement rate (EER) on dynamic contrast-enhanced (DCE)-MRI,¹³² can accompany pain reduction following steroid injection. MRI evidence of BMLs has been used to select patients for recruitment to trials of bisphosphonates, primarily on the basis that BMLs predict structural progression. Initial trials were designed to reduce structural progression by inhibiting subchondral osteoclast activity. Reductions in BMLs were accompanied by reductions in pain following intravenous zoledronate³⁶ or neridronate.³⁷ It remains unclear whether bisphosphonates would have demonstrated similar analgesic benefit in people without BMLs, although earlier RCTs of bisphosphonates in unselected patients with OA showed less consistent analgesic benefits.¹³³

Toward Patient Stratification

Biomarkers have potential to stratify patients, identifying those most likely to benefit from each of a range of treatments. However, further research is needed to determine which specific marker might best, and independently, predict treatment outcomes. Diverse biomarkers correlate with each other, and biomarker combinations might prove more specific and sensitive for detecting patient subgroups. For example, patients with a predominantly peripheral drive to their OA pain might display combinations of BMLs, synovitis, and loss of osteochondral integrity, while those with a major contribution for alterations in central pain processing might display changes in quantitative sensory tests, negative affect, and diurnal fatigue. Such biomarker combinations might cross traditional disciplinary boundaries, and it is currently little understood how wet biomarkers might correspond to imaging evidence of OA disease activity or, in combination, might increase their predictive value.¹³⁴ Some blood biomarkers of inflammation display only weak correlation with imaging evidence of synovitis by ultrasound or MRI, although it remains possible that more specific circulating cytokines might inform of the nature of imaging-determined synovitis, and thereby indicate specific treatment targets.

Patient stratification depends on a degree of phenotypic stability over a period of therapy. Defining phenotypes that remain stable over long periods approaches diagnostic classification. It is currently unclear whether synovitis reflects a stable OA phenotype or fluctuates throughout the disease. Latent class analysis of pathological samples has suggested

that OA knees can be grouped largely according to the intensity of histological synovitis (scored based on hyperplasia and lymphoid infiltration), and that such groups are concealed among patients with similar clinical characteristics and demographics.³ Generalized nodal OA has discrete genetic risk factors¹³⁵ and displays moderate synovitis,¹³⁶ further suggesting that inflammatory OA subgroups might be contained within the wider OA population. Synovitis has been identified in both early and established OA, and association with articular damage indicates a risk factor for structural progression, rather than a characteristic restricted to late-stage disease.¹³⁷ Synovitis might also be episodic, and BMLs are dynamic changes in OA, which might fluctuate from week to week.¹³⁸ Increases and decreases in synovitis or BML size correspond to changing OA pain, suggesting that pain might be an indicator of OA inflammatory or subchondral “disease activity.” Offering treatments during periods of active disease might be more acceptable to patients who might experience protracted periods of symptomatic remission, and might avoid risks from unnecessary treatment. Accessible and acceptable biomarkers that can define inflammatory, osteochondral, or neuronal mechanisms that underlie active disease could inform the management of OA flares.

OA biomarker studies remain in their infancy, having progressed from investigating biomarkers of structural disease to the clinically pertinent issue of OA pain. Key areas for future research will be to define those biomarkers alone or in combination that best predict the need for treatment or successful treatment outcome. RCTs of novel treatments aiming to benefit a subgroup of patients with OA will need to be paired with suitable biomarkers that will permit cost-effective treatment in clinical practice. Although biomarkers of structural disease might also predict pain outcomes, other biomarkers might await discovery that better predict pain than structural disease.

Summary

OA pain remains a major source of distress and disability, and is expected to increase as our populations continue to age. Current pharmacological analgesics have limited efficacy for what is typically a long-term condition, and their use is often restricted by clinically important adverse events. Our increasing understanding of the complex mechanisms that underlie OA pain offers a wide range of potential new treatment targets, and expectation is high that improved treatments will be available in the near future. New drugs for OA pain might come from repurposing those developed for other conditions (depression, anxiety, neuropathic pain, osteoporosis, and inflammatory arthritis), as well as novel compounds targeting pain mechanisms specific to the joint. Medication is unlikely to be a complete solution for people with OA, and even when pain relief is achieved, non-medical interventions, such as physiotherapy, occupational

therapy, orthotics, or psychological treatment, might be needed to restore normal function and well-being. Total joint replacement is one of the most effective treatments available in modern medicine for improving quality of life, and yet many people do not return to previously valued activities. New treatments are needed, and their development requires careful evaluation of their benefits and possible adverse events, within the context of the key needs of people with OA. A silver bullet is unlikely for such a heterogenous disease as OA, but new treatments will extend the benefit of existing therapies, while not necessarily replacing them all.

Abbreviations

| | |
|-----------|--|
| 2-AG | 2-arachidonoylglycerol |
| ADAMTS5 | A disintegrin and metalloproteinase with thrombospondin motifs 5 |
| AMPA/KA | Ampa kainate subtype of the glutamate receptor |
| ASIC2 | Acid-sensing ion channel 2 |
| BDNF | Brain-derived neurotrophic factor |
| BMI | Body mass index |
| BMLs | Bone marrow lesions |
| BMP | Bone morphogenetic protein |
| CB1R/CB2R | Cannabinoid receptors 1 and 2 |
| CCL | C-C Motif Chemokine Ligand |
| CCDC | Coiled-coil domain containing |
| CD14 | Cluster of differentiation 14 |
| CDK | Cyclin-dependent kinase |
| CGRP | Calcitonin gene-related peptide |
| COMP | Cartilage oligomeric matrix protein |
| COMT | Catecholamine O-methyl transferase |
| CPM | Conditioned pain modulation |
| CRAC | Calcium-release activated calcium |
| CRP | C-reactive protein |
| CSF | Cerebrospinal fluid |
| CX3CL1 | Fractalkine |
| DCE | Dynamic contrast-enhanced |
| DRG | Dorsal root ganglion |
| EER | Early enhancement rate |
| EGF | Epidermal growth factor |
| ESR | Erythrocyte sedimentation rate |
| Fas | Tumour necrosis factor receptor superfamily, member 6 |
| FGF | Fibroblast growth factor |
| Flt3 | Fms-related Tyrosine Kinase 3 |
| Fms | Feline McDonough Sarcoma |
| GM-CSF | Granulocyte-macrophage colony-stimulating factor |
| HMGB | High mobility group box |
| ICE | Interleukin-1 β converting enzyme |
| IFN | Interferon |
| IK 1 | Inward rectifier current |
| IL | Interleukin |
| IL-1-RA | Interleukin 1 receptor antagonist |
| Kit | Stem cell factor receptor |

| | |
|---------|--|
| LNc RNA | Long noncoding RNA |
| MMP | Matrix metalloproteinases |
| MTF1 | Metalregulatory transcription factor-1 |
| MRI | Magnetic resonance imaging |
| NGF | Nerve growth factor |
| NMDA | N-Methyl-D-aspartic acid |
| NSAIDs | Nonsteroidal anti-inflammatory drugs |
| OA | Osteoarthritis |
| PACE4 | Paired amino acid converting enzyme 4 |
| PBL | Peripheral Blood Leucocytes |
| PEDF | Pigment epithelium-derived factor |
| PGE | Prostaglandin E |
| RARB | Retinoic Acid Receptor Beta |
| RCT | Randomised controlled trial |
| RGD | Arg-Gly-Asp |
| RPOA | Rapidly progressive osteoarthritis |
| RA | Rheumatoid arthritis |
| sHA | Serum hyaluronic acid |
| SSNRI | Selective serotonin/norepinephrine reuptake inhibitor |
| sTNFR | Soluble tumour necrosis factor receptor |
| TGF | Transforming growth factor |
| TIMP | Tissue inhibitor of metalloproteinases |
| TMCO3 | Transmembrane and Coiled-Coil Domains 3 |
| TNF | Tumor Necrosis Factor |
| TPV1 | Transient receptor potential cation channel subfamily V member 1 |
| TRAP5b | Tartrate resistant acid phosphatase type b5 |
| Trk | Tropomyosin receptor kinase |
| TRPC3 | Short transient receptor potential channel 3 |
| TSPAN | Tetraspanin |
| TSPO | Translocator protein |
| TPV1 | Transient receptor potential cation channel subfamily V member 1 |
| uCTX | Urinary crosslinked C-telopeptide |
| VEGF | Vascular endothelial growth factor |
| YKL-40 | Chitinase-3-like protein 1 (CHI3L1) |

Acknowledgments

The authors gratefully acknowledge Mr Tom Kurien and Dr Aliya Sarmanova for the radiographic, MRI, and ultrasound images.

Declaration of Conflicting Interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

References

1. Altman, R.; Asch, E.; Bloch, D.; et al. Development of Criteria for the Classification and Reporting of Osteoarthritis. Classification of Osteoarthritis of the Knee. Diagnostic and

- Therapeutic Criteria Committee of the American Rheumatism Association. *Arthritis Rheum.* **1986**, *29* (8), 1039–1049.
2. Bastick, A. N.; Belo, J. N.; Runhaar, J.; et al. What Are the Prognostic Factors for Radiographic Progression of Knee Osteoarthritis? A Meta-Analysis. *Clin. Orthop.* **2015**, *473* (9), 2969–2989.
 3. Wyatt, L. A.; Moreton, B. J.; Mapp, P. I.; et al. Histopathological Subgroups in Knee Osteoarthritis. *Osteoarthr. Cartil.* **2017**, *25* (1), 14–22.
 4. Walsh, D. A.; McWilliams, D. F. Pain in Rheumatoid Arthritis. *Curr. Pain Headache Rep.* **2012**, *16* (6), 509–517.
 5. Makris, E. A.; Gomoll, A. H.; Malizos, K. N.; et al. Repair and Tissue Engineering Techniques for Articular Cartilage. *Nat. Rev. Rheumatol.* **2015**, *11* (1), 21–34.
 6. Raastad, J.; Reiman, M.; Coeytaux, R.; et al. The Association between Lumbar Spine Radiographic Features and Low Back Pain: A Systematic Review and Meta-Analysis. *Semin. Arthritis Rheum.* **2015**, *44* (5), 571–585.
 7. Maas, E. T.; Ostelo, R. W.; Niemisto, L.; et al. Radiofrequency Denervation for Chronic Low Back Pain. *Cochrane Database Syst. Rev.* **2015**, *10*, CD008572.
 8. Culvenor, A. G.; Wirth, W.; Roth, M.; et al. Predictive Capacity of Thigh Muscle Strength in Symptomatic and/or Radiographic Knee Osteoarthritis Progression: Data from the Foundation for the National Institutes of Health Osteoarthritis Biomarkers Consortium. *Am. J. Phys. Med. Rehabil.* **2016**, *95* (12), 931–938.
 9. Roddy, E.; Zhang, W.; Doherty, M. Aerobic Walking or Strengthening Exercise for Osteoarthritis of the Knee? A Systematic Review. *Ann. Rheum. Dis.* **2005**, *64* (4), 544–548.
 10. Park, D. Y.; Min, B. H.; Kim, D. W.; et al. Polyethylene Wear Particles Play a Role in Development of Osteoarthritis via Detrimental Effects on Cartilage, Meniscus, and Synovium. *Osteoarthr. Cartil.* **2013**, *21* (12), 2021–2029.
 11. Xu, L.; Hayashi, D.; Roemer, F. W.; et al. Magnetic Resonance Imaging of Subchondral Bone Marrow Lesions in Association with Osteoarthritis. *Semin Arthritis Rheum* **2012**, *42* (2), 105–118.
 12. Walsh, D. A.; McWilliams, D. F.; Turley, M. J.; et al. Angiogenesis and Nerve Growth Factor at the Osteochondral Junction in Rheumatoid Arthritis and Osteoarthritis. *Rheumatology (Oxford)* **2010**, *49* (10), 1852–1861.
 13. Hall, M.; Doherty, S.; Courtney, P.; et al. Synovial Pathology Detected on Ultrasound Correlates with the Severity of Radiographic Knee Osteoarthritis More Than with Symptoms. *Osteoarthr. Cartil.* **2014**, *22* (10), 1627–1633.
 14. *Osteoarthritis: Care and Management in Adults*, NICE Clinical Guideline 177; National Institute for Health and Care Excellence: London, **2014**.
 15. Dakin, H.; Gray, A.; Fitzpatrick, R.; et al. Rationing of Total Knee Replacement: A Cost-Effectiveness Analysis on a Large Trial Data Set. *BMJ Open* **2012**, *2* (1), e000332.
 16. Suokas, A. K.; Sagar, D. R.; Mapp, P. I.; et al. Design, Study Quality and Evidence of Analgesic Efficacy in Studies of Drugs in Models of OA Pain: A Systematic Review and a Meta-Analysis. *Osteoarthr. Cartil.* **2014**, *22* (9), 1207–1223.
 17. Berenbaum, F. Osteoarthritis as an Inflammatory Disease (Osteoarthritis Is Not Osteoarthrosis!). *Osteoarthr. Cartil.* **2013**, *21* (1), 16–21.
 18. Tsuchida, A. I.; Beekhuizen, M.; 't Hart, M. C.; et al. Cytokine Profiles in the Joint Depend on Pathology, but Are Different between Synovial Fluid, Cartilage Tissue and Cultured Chondrocytes. *Arthritis Res. Ther.* **2014**, *16* (5), 441.
 19. Egg, D. Concentrations of prostaglandins D2, E2, F2 Alpha, 6-keto-F1 Alpha and Thromboxane B2 in Synovial Fluid from Patients with Inflammatory Joint Disorders and Osteoarthritis. *Z. Rheumatol.* **1984**, *43* (2), 89–96.
 20. Jin, X.; Beguerie, J. R.; Zhang, W.; et al. Circulating C Reactive Protein in Osteoarthritis: A Systematic Review and Meta-Analysis. *Ann. Rheum. Dis.* **2015**, *74* (4), 703–710.
 21. Nelson, A. E.; Allen, K. D.; Golightly, Y. M.; et al. A Systematic Review of Recommendations and Guidelines for the Management of Osteoarthritis: The Chronic Osteoarthritis Management Initiative of the U.S. Bone and Joint Initiative. *Semin. Arthritis Rheum.* **2014**, *43* (6), 701–712.
 22. Wenham, C. Y.; Hensor, E. M.; Grainger, A. J.; et al. A Randomized, Double-Blind, Placebo-Controlled Trial of Low-Dose Oral Prednisolone for Treating Painful Hand Osteoarthritis. *Rheumatology (Oxford)* **2012**, *51* (12), 2286–2294.
 23. Wenham, C. Y.; Grainger, A. J.; Hensor, E. M.; et al. Methotrexate for Pain Relief in Knee Osteoarthritis: An Open-Label Study. *Rheumatology (Oxford)* **2013**, *52* (5), 888–892.
 24. Kingsbury, S. R.; Tharmanathan, P.; Adamson, J.; et al. Hydroxychloroquine Effectiveness in Reducing Symptoms of Hand Osteoarthritis (HERO): Study Protocol for a Randomized Controlled Trial. *Trials* **2013**, *14*, 64.
 25. Kingsbury, S. R.; Tharmanathan, P.; Arden, N. K.; et al. Pain Reduction with Oral Methotrexate in Knee Osteoarthritis, a Pragmatic Phase III Trial of Treatment Effectiveness (PROMOTE): Study Protocol for a Randomized Controlled Trial. *Trials* **2015**, *16*, 77.
 26. Detert, J.; Klaus, P.; Listing, J.; et al. Hydroxychloroquine in Patients with Inflammatory and Erosive Osteoarthritis of the Hands (OA TREAT): Study Protocol for a Randomized Controlled Trial. *Trials* **2014**, *15*, 412.
 27. Walsh, D. A.; McWilliams, D. F. Mechanisms, Impact and Management of Pain in Rheumatoid Arthritis. *Nat. Rev. Rheumatol.* **2014**, *10* (10), 581–592.
 28. Chevalier, X.; Ravaut, P.; Maheu, E.; et al. Adalimumab in Patients with Hand Osteoarthritis Refractory to Analgesics and NSAIDs: A Randomised, Multicentre, Double-Blind, Placebo-Controlled Trial. *Ann. Rheum. Dis.* **2015**, *74* (9), 1697–1705.
 29. Verbruggen, G.; Wittoek, R.; Vander Cruyssen, B.; et al. Tumour Necrosis Factor Blockade for the Treatment of Erosive Osteoarthritis of the Interphalangeal Finger Joints: A Double Blind, Randomised Trial on Structure Modification. *Ann. Rheum. Dis.* **2012**, *71* (6), 891–898.
 30. Grunke, M.; Schulze-Koops, H. Successful Treatment of Inflammatory Knee Osteoarthritis with Tumour Necrosis Factor Blockade. *Ann. Rheum. Dis.* **2006**, *65* (4), 555–556.
 31. Zanetti, M.; Bruder, E.; Romero, J.; et al. Bone Marrow Edema Pattern in Osteoarthritic Knees: Correlation between MR Imaging and Histologic Findings. *Radiology* **2000**, *215* (3), 835–840.
 32. Prieto-Potin, I.; Largo, R.; Roman-Blas, J. A.; et al. Characterization of Multinucleated Giant Cells in Synovium

- and Subchondral Bone in Knee Osteoarthritis and Rheumatoid Arthritis. *BMC Musculoskelet. Disord.* **2015**, *16*, 226.
33. Sagar, D. R.; Ashraf, S.; Xu, L.; et al. Osteoprotegerin Reduces the Development of Pain Behaviour and Joint Pathology in a Model of Osteoarthritis. *Ann. Rheum. Dis.* **2014**, *73* (8), 1558–1565.
 34. Strassle, B. W.; Mark, L.; Leventhal, L.; et al. Inhibition of Osteoclasts Prevents Cartilage Loss and Pain in a Rat Model of Degenerative Joint Disease. *Osteoarthr. Cartil.* **2010**, *18* (10), 1319–1328.
 35. McDougall, J. J.; Schuelert, N.; Bowyer, J. Cathepsin K Inhibition Reduces CTXII Levels and Joint Pain in the Guinea Pig Model of Spontaneous Osteoarthritis. *Osteoarthr. Cartil.* **2010**, *18* (10), 1355–1357.
 36. Laslett, L. L.; Dore, D. A.; Quinn, S. J.; et al. Zoledronic Acid Reduces Knee Pain and Bone Marrow Lesions over 1 Year: A Randomised Controlled Trial. *Ann. Rheum. Dis.* **2012**, *71* (8), 1322–1328.
 37. Varenna, M.; Zucchi, F.; Failoni, S.; et al. Intravenous Neridronate in the Treatment of Acute Painful Knee Osteoarthritis: A Randomized Controlled Study. *Rheumatology (Oxford)* **2015**, *54* (10), 1826–1832.
 38. Nishii, T.; Tamura, S.; Shiomi, T.; et al. Alendronate Treatment for Hip Osteoarthritis: Prospective Randomized 2-Year Trial. *Clin. Rheumatol.* **2013**, *32* (12), 1759–1766.
 39. Pelletier, J. P.; Roubille, C.; Raynauld, J. P.; et al. Disease-Modifying Effect of Strontium Ranelate in a Subset of Patients from the Phase III Knee Osteoarthritis Study SEKIOA Using Quantitative MRI: Reduction in Bone Marrow Lesions Protects against Cartilage Loss. *Ann. Rheum. Dis.* **2015**, *74* (2), 422–429.
 40. Reginster, J. Y.; Badurski, J.; Bellamy, N.; et al. Efficacy and Safety of Strontium Ranelate in the Treatment of Knee Osteoarthritis: Results of a Double-Blind, Randomised Placebo-Controlled Trial. *Ann. Rheum. Dis.* **2013**, *72* (2), 179–186.
 41. Hawker, G. A.; Stewart, L.; French, M. R.; et al. Understanding the Pain Experience in Hip and Knee Osteoarthritis—An OARSI/OMERACT Initiative. *Osteoarthr. Cartil.* **2008**, *16* (4), 415–422.
 42. Moreton, B. J.; Wheeler, M.; Walsh, D. A.; et al. Rasch Analysis of the Intermittent and Constant Osteoarthritis Pain (ICOAP) Scale. *Osteoarthr. Cartil.* **2012**, *20* (10), 1109–1115.
 43. McGaraghty, S.; Chu, K. L.; Perner, R. J.; et al. TRPA1 Modulation of Spontaneous and Mechanically Evoked Firing of Spinal Neurons in Uninjured, Osteoarthritic, and Inflamed Rats. *Mol. Pain.* **2010**, *6*, 14.
 44. Quick, K.; Zhao, J.; Eijkelkamp, N.; et al. TRPC3 and TRPC6 Are Essential for Normal Mechanotransduction in Subsets of Sensory Neurons and Cochlear Hair Cells. *Open Biol.* **2012**, *2* (5), 120068.
 45. Eijkelkamp, N.; Linley, J. E.; Torres, J. M.; et al. A Role for Piezo2 in EPAC1-Dependent Mechanical Allodynia. *Nat. Commun.* **2013**, *4*, 1682.
 46. Wood, J. N.; Eijkelkamp, N. Noxious Mechanosensation—Molecules and Circuits. *Curr. Opin. Pharmacol.* **2012**, *12* (1), 4–8.
 47. Skou, S. T.; Roos, E. M.; Simonsen, O.; et al. The Efficacy of Non-Surgical Treatment on Pain and Sensitization in Patients with Knee Osteoarthritis: A Pre-Defined Ancillary Analysis from a Randomized Controlled Trial. *Osteoarthr. Cartil.* **2016**, *24* (1), 108–116.
 48. Brouwer, R. W.; Huizinga, M. R.; Duivenvoorden, T.; et al. Osteotomy for Treating Knee Osteoarthritis. *Cochrane Database Syst. Rev.* **2014**, *12*, CD004019.
 49. Lu, Y.; Ma, X.; Sabharwal, R.; et al. The Ion Channel ASIC2 Is Required for Baroreceptor and Autonomic Control of the Circulation. *Neuron* **2009**, *64* (6), 885–897.
 50. Creamer, P.; Hunt, M.; Dieppe, P. Pain Mechanisms in Osteoarthritis of the Knee: Effect of Intraarticular Anesthetic. *J. Rheumatol.* **1996**, *23* (6), 1031–1036.
 51. Hassan, B. S.; Doherty, S. A.; Mockett, S.; et al. Effect of Pain Reduction on Postural Sway, Proprioception, and Quadriceps Strength in Subjects with Knee Osteoarthritis. *Ann. Rheum. Dis.* **2002**, *61* (5), 422–428.
 52. Rahman, W.; Dickenson, A. H. Osteoarthritis-Dependent Changes in Antinociceptive Action of Nav1.7 and Nav1.8 Sodium Channel Blockers: An In Vivo Electrophysiological Study in the Rat. *Neuroscience* **2015**, *295*, 103–116.
 53. Schnitzer, T. J.; Marks, J. A. A Systematic Review of the Efficacy and General Safety of Antibodies to NGF in the Treatment of OA of the Hip or Knee. *Osteoarthr. Cartil.* **2015**, *23* (Suppl. 1), S8–S17.
 54. Gow, J. M.; Tsuji, W. H.; Williams, G. J.; et al. Safety, Tolerability, Pharmacokinetics, and Efficacy of AMG 403, a Human Anti-Nerve Growth Factor Monoclonal Antibody, in Two Phase I Studies with Healthy Volunteers and Knee Osteoarthritis Subjects. *Arthritis Res. Ther.* **2015**, *17*, 282.
 55. Katz, N.; Borenstein, D. G.; Birbara, C.; et al. Efficacy and Safety of Tanezumab in the Treatment of Chronic Low Back Pain. *Pain* **2011**, *152* (10), 2248–2258.
 56. Nwosu, L. N.; Mapp, P. I.; Chapman, V.; et al. Blocking the Tropomyosin Receptor Kinase A (TrkA) Receptor Inhibits Pain Behaviour in Two Rat Models of Osteoarthritis. *Ann. Rheum. Dis.* **2016**, *75* (6), 1246–1254.
 57. Xu, L.; Nwosu, L. N.; Mapp, P. I.; et al. The Anti-NGF Antibody muMab 911 Both Prevents and Reverses Pain Behaviour and Subchondral Osteoclast Numbers in a Rat Model of Osteoarthritis Pain. *Osteoarthr. Cartil.* **2016**, *24* (9), 1587–1595.
 58. Mapp, P. I.; Walsh, D. A. Mechanisms and Targets of Angiogenesis and Nerve Growth in Osteoarthritis. *Nat. Rev. Rheumatol.* **2012**, *8* (7), 390–398.
 59. Bullock, C. M.; Wookey, P.; Bennett, A.; et al. Peripheral Calcitonin Gene-Related Peptide Receptor Activation and Mechanical Sensitization of the Joint in Rat Models of Osteoarthritis Pain. *Arthritis Rheumatol.* **2014**, *66* (8), 2188–2200.
 60. Walsh, D. A.; Mapp, P. I.; Kelly, S. Calcitonin Gene-Related Peptide in the Joint: Contributions to Pain and Inflammation. *Br. J. Clin. Pharmacol.* **2015**, *80* (5), 965–978.
 61. Arendt-Nielsen, L. Central Sensitization in Humans: Assessment and Pharmacology. *Handb. Exp. Pharmacol.* **2015**, *227*, 79–102.
 62. Zhou, H. Y.; Chen, S. R.; Pan, H. L. Targeting N-Methyl-D-Aspartate Receptors for Treatment of Neuropathic Pain. *Expert Rev. Clin. Pharmacol.* **2011**, *4* (3), 379–388.
 63. Lindstedt, F.; Berrebi, J.; Greayer, E.; et al. Conditioned Pain Modulation Is Associated with Common Polymorphisms in the Serotonin Transporter Gene. *PLoS One* **2011**, *6* (3), e18252.
 64. Treister, R.; Pud, D.; Ebstein, R. P.; et al. Association between Polymorphisms in Serotonin and Dopamine-

- Related Genes and endogenous Pain Modulation. *J. Pain* **2011**, *12* (8), 875–883.
65. Yarnitsky, D.; Granot, M.; Nahman-Averbuch, H.; et al. Conditioned Pain Modulation Predicts Duloxetine Efficacy in Painful Diabetic Neuropathy. *Pain* **2012**, *153* (6), 1193–1198.
 66. Clair, A.; Emir, B. The Safety and Efficacy of Pregabalin for Treating Subjects with Fibromyalgia and Moderate or Severe Baseline Widespread Pain. *Curr. Med. Res. Opin.* **2016**, *32* (3), 601–609.
 67. Jaffe, M.; Young, J. D. I. J. P.; Spiegel, K.; et al. Post-Hoc Results Show Beneficial Effects of Pregabalin in Patients with Osteoarthritis of the Hip. *Arthritis Rheum.* **2000**, S337.
 68. Wang, Z. Y.; Shi, S. Y.; Li, S. J.; et al. Efficacy and Safety of Duloxetine on Osteoarthritis Knee Pain: A Meta-Analysis of Randomized Controlled Trials. *Pain Med.* **2015**, *16* (7), 1373–1385.
 69. Zhang, W.; Robertson, J.; Jones, A. C.; et al. The Placebo Effect and Its Determinants in Osteoarthritis: Meta-Analysis of Randomised Controlled Trials. *Ann. Rheum. Dis.* **2008**, *67* (12), 1716–1723.
 70. Axford, J.; Butt, A.; Heron, C.; et al. Prevalence of Anxiety and Depression in Osteoarthritis: Use of the Hospital Anxiety and Depression Scale as a Screening Tool. *Clin. Rheumatol.* **2010**, *29* (11), 1277–1283.
 71. Baliki, M. N.; Geha, P. Y.; Jabakhanji, R.; et al. A Preliminary fMRI Study of Analgesic Treatment in Chronic Back Pain and Knee Osteoarthritis. *Mol. Pain* **2008**, *4*, 47.
 72. Mochcovitch, M. D.; da Rocha Freire, R. C.; Garcia, R. F.; et al. A Systematic Review of fMRI Studies in Generalized Anxiety Disorder: Evaluating Its Neural and Cognitive Basis. *J. Affect. Disord.* **2014**, *167*, 336–342.
 73. Behbehani, M. M. Functional Characteristics of the Midbrain Periaqueductal Gray. *Progr. Neurobiol.* **1995**, *46* (6), 575–605.
 74. de Souza, J. B.; Potvin, S.; Goffaux, P.; et al. The Deficit of Pain Inhibition in Fibromyalgia Is More Pronounced in Patients with Comorbid Depressive Symptoms. *Clin. J. Pain* **2009**, *25* (2), 123–127.
 75. Vidor, L. P.; Torres, I. L.; Medeiros, L. F.; et al. Association of Anxiety with Intracortical Inhibition and Descending Pain Modulation in Chronic Myofascial Pain Syndrome. *BMC Neurosci.* **2014**, *15*, 42.
 76. Goodin, B. R.; Glover, T. L.; Sotolongo, A.; et al. The Association of Greater Dispositional Optimism with Less Endogenous Pain Facilitation Is Indirectly Transmitted through Lower Levels of Pain Catastrophizing. *J. Pain* **2013**, *14* (2), 126–135.
 77. Thompson, T.; Keogh, E.; French, C. C.; et al. Anxiety Sensitivity and Pain: Generalisability across Noxious Stimuli. *Pain* **2008**, *134* (1–2), 187–196.
 78. Ang, D. C.; Chakr, R.; Mazzuca, S.; et al. Cognitive-Behavioral Therapy Attenuates Nociceptive Responding in Patients with Fibromyalgia: A Pilot Study. *Arthritis Care Res. (Hoboken)* **2010**, *62* (5), 618–623.
 79. Dickens, C.; McGowan, L.; Dale, S. Impact of Depression on Experimental Pain Perception: A Systematic Review of the Literature with Meta-Analysis. *Psychosom. Med.* **2003**, *65* (3), 369–375.
 80. Sullivan, M.; Tanzer, M.; Stanish, W.; et al. Psychological Determinants of Problematic Outcomes Following Total Knee Arthroplasty. *Pain* **2009**, *143* (1–2), 123–129.
 81. Wertli, M. M.; Rasmussen-Barr, E.; Held, U.; et al. Fear-Avoidance Beliefs—A Moderator of Treatment Efficacy in Patients with Low Back Pain: A Systematic Review. *Spine J.* **2014**, *14* (11), 2658–2678.
 82. Schaible, H. G. Nociceptive Neurons Detect Cytokines in Arthritis. *Arthritis Res. Ther.* **2014**, *16* (5), 470.
 83. Graven-Nielsen, T.; Wodehouse, T.; Langford, R. M.; et al. Normalization of Widespread Hyperesthesia and Facilitated Spatial Summation of Deep-Tissue Pain in Knee Osteoarthritis Patients after Knee Replacement. *Arthritis Rheum.* **2012**, *64* (9), 2907–2916.
 84. Kosek, E.; Ordeberg, G. Lack of Pressure Pain Modulation by Heterotopic Noxious Conditioning Stimulation in Patients with Painful Osteoarthritis Before, but Not Following, Surgical Pain Relief. *Pain* **2000**, *88* (1), 69–78.
 85. Martinez-Jauand, M.; Sitges, C.; Rodriguez, V.; et al. Pain Sensitivity in Fibromyalgia Is Associated with Catechol-O-Methyltransferase (COMT) Gene. *Eur. J. Pain* **2013**, *17* (1), 16–27.
 86. Sato, H.; Droney, J.; Ross, J.; et al. Gender, Variation in Opioid Receptor Genes and Sensitivity to Experimental Pain. *Mol. Pain* **2013**, *9*, 20.
 87. Petersen, K. K.; Arendt-Nielsen, L.; Simonsen, O.; et al. Presurgical Assessment of Temporal Summation of Pain Predicts the Development of Chronic Postoperative Pain 12 Months after Total Knee Replacement. *Pain* **2015**, *156* (1), 55–61.
 88. Neogi, T.; Guermazi, A.; Roemer, F.; et al. Association of Joint Inflammation with Pain Sensitization in Knee Osteoarthritis: The Multicenter Osteoarthritis Study. *Arthritis Rheumatol.* **2016**, *68* (3), 654–661.
 89. Arendt-Nielsen, L.; Eskehave, T. N.; Egsgaard, L. L.; et al. Association between Experimental Pain Biomarkers and Serologic Markers in Patients with Different Degrees of Painful Knee Osteoarthritis. *Arthritis Rheumatol.* **2014**, *66* (12), 3317–3326.
 90. Lee, Y. C.; Lu, B.; Bathon, J. M.; et al. Pain Sensitivity and Pain Reactivity in Osteoarthritis. *Arthritis Care Res. (Hoboken)* **2011**, *63* (3), 320–327.
 91. Mapp, P. I.; Sagar, D. R.; Ashraf, S.; et al. Differences in Structural and Pain Phenotypes in the Sodium Monoiodoacetate and Meniscal Transection Models of Osteoarthritis. *Osteoarthritis Cartil.* **2013**, *21* (9), 1336–1345.
 92. Mannion, R. J.; Costigan, M.; Decosterd, I.; et al. Neurotrophins: Peripherally and Centrally Acting Modulators of Tactile Stimulus-Induced Inflammatory Pain Hypersensitivity. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96* (16), 9385–9390.
 93. Sagar, D. R.; Burston, J. J.; Hathway, G. J.; et al. The Contribution of Spinal Glial Cells to chronic Pain Behaviour in the Monosodium Iodoacetate Model of Osteoarthritic Pain. *Mol. Pain* **2011**, *7*, 88.
 94. Loggia, M. L.; Chonde, D. B.; Akeju, O.; et al. Evidence for Brain Glial Activation in Chronic Pain Patients. *Brain* **2015**, *138* (Pt 3), 604–615.
 95. Kosek, E.; Altawil, R.; Kadetoff, D.; et al. Evidence of Different Mediators of Central Inflammation in Dysfunctional and Inflammatory Pain—Interleukin-8 in Fibromyalgia and Interleukin-1 Beta in Rheumatoid Arthritis. *J. Neuroimmunol.* **2015**, *280*, 49–55.
 96. Guo, S. L.; Han, C. T.; Jung, J. L.; et al. Cystatin C in Cerebrospinal Fluid Is Upregulated in Elderly Patients with

- Chronic Osteoarthritis Pain and Modulated through Matrix Metalloproteinase 9-Specific Pathway. *Clin. J. Pain* **2014**, *30* (4), 331–339.
97. Zhang, W.; Nuki, G.; Moskowitz, R. W.; et al. OARSI Recommendations for the Management of Hip and Knee Osteoarthritis: Part III: Changes in Evidence Following Systematic Cumulative Update of Research Published through January 2009. *Osteoarthr. Cartil.* **2010**, *18* (4), 476–499.
98. Cimmino, M. A.; Scarpa, R.; Caporali, R.; et al. Body Mass and Osteoarthritic Pain: Results from a Study in General Practice. *Clin. Exp. Rheumatol.* **2013**, *31* (6), 843–849.
99. Kim, C. H.; Luedtke, C. A.; Vincent, A.; et al. Association of Body Mass Index with Symptom Severity and Quality of Life in Patients with Fibromyalgia. *Arthritis Care Res. (Hoboken)* **2012**, *64* (2), 222–228.
100. Kutlu, S.; Canpolat, S.; Sandal, S.; et al. Effects of Central and Peripheral Administration of Leptin on Pain Threshold in Rats and Mice. *Neuroendocrinol. Lett.* **2003**, *24* (3–4), 193–196.
101. Iannitti, T.; Graham, A.; Dolan, S. Increased Central and Peripheral Inflammation and Inflammatory Hyperalgesia in Zucker Rat Model of Leptin Receptor Deficiency and Genetic Obesity. *Exp. Physiol.* **2012**, *97* (11), 1236–1245.
102. Pincus, T.; Holt, N.; Vogel, S.; et al. Cognitive and Affective Reassurance and Patient Outcomes in Primary Care: A Systematic Review. *Pain* **2013**, *154* (11), 2407–2416.
103. Farquhar-Smith, P.; Gubbay, A. Tramadol and Acetaminophen Combination for Chronic Non-Cancer Pain. *Expert Opin. Pharmacother.* **2013**, *14* (16), 2297–2304.
104. Roberts, E.; Delgado Nunes, V.; Buckner, S.; et al. Paracetamol: Not as Safe as We Thought? A Systematic Literature Review of Observational Studies. *Ann. Rheum. Dis.* **2016**, *75* (3), 552–559.
105. Hochberg, M. C.; Tive, L. A.; Abramson, S. B.; et al. When Is Osteonecrosis Not Osteonecrosis? Adjudication of Reported Serious Adverse Joint Events in the Tanezumab Clinical Development Program. *Arthritis Rheumatol.* **2016**, *68* (2), 382–391.
106. Schnitzer, T. J.; Ekman, E. F.; Spierings, E. L.; et al. Efficacy and Safety of Tanezumab Monotherapy or Combined with Non-Steroidal Anti-Inflammatory Drugs in the Treatment of Knee or Hip Osteoarthritis Pain. *Ann. Rheum. Dis.* **2015**, *74* (6), 1202–1211.
107. LaBranche, T. P.; Bendele, A. M.; Omura, B. C.; et al. Nerve Growth Factor Inhibition with Tanezumab Influences Weight-Bearing and Subsequent Cartilage Damage in the Rat Medial Meniscal Tear Model. *Ann. Rheum. Dis.* **2017**, *76* (1), 295–302.
108. Tesfaye, S.; Wilhelm, S.; Lledo, A.; et al. Duloxetine and Pregabalin: High-Dose Monotherapy or Their Combination? The “COMBO-DN Study”—A Multinational, Randomized, Double-Blind, Parallel-Group Study in Patients with Diabetic Peripheral Neuropathic Pain. *Pain* **2013**, *154* (12), 2616–2625.
109. Valdes, A. M.; Abhishek, A.; Muir, K.; et al. Association of Beta-Blocker Use with Less Prevalent Joint Pain and Lower Opioid Requirement in People with Osteoarthritis. *Arthritis Care Res. (Hoboken)* **2016**. DOI: 10.1002/acr.23091.
110. Trijau, S.; Avouac, J.; Escalas, C.; et al. Influence of Flare Design on Symptomatic Efficacy of Non-Steroidal Anti-Inflammatory Drugs in Osteoarthritis: A Meta-Analysis of Randomized Placebo-Controlled Trials. *Osteoarthr. Cartil.* **2010**, *18* (8), 1012–1018.
111. Zeidan, F.; Lobanov, O. V.; Kraft, R. A.; et al. Brain Mechanisms Supporting Violated Expectations of Pain. *Pain* **2015**, *156* (9), 1772–1785.
112. Bauer, D. C.; Hunter, D. J.; Abramson, S. B.; et al. Classification of Osteoarthritis Biomarkers: A Proposed Approach. *Osteoarthr. Cartil.* **2006**, *14* (8), 723–727.
113. Kraus, V. B.; Collins, J. E.; Hargrove, D.; et al. Predictive Validity of Biochemical Biomarkers in Knee Osteoarthritis: Data from the FNIH OA Biomarkers Consortium. *Ann. Rheum. Dis.* **2016**, *76* (1), 186–195.
114. Kluzek, S.; Bay-Jensen, A. C.; Judge, A.; et al. Serum Cartilage Oligomeric Matrix Protein and Development of Radiographic and Painful Knee Osteoarthritis. A Community-Based Cohort of Middle-Aged Women. *Biomarkers* **2015**, *20* (8), 557–564.
115. Zou, K.; Wong, J.; Abdullah, N.; et al. Examination of Overall Treatment Effect and the Proportion Attributable to Contextual Effect in Osteoarthritis: Meta-Analysis of Randomised Controlled Trials. *Ann. Rheum. Dis.* **2016**, *75* (11), 1964–1970.
116. Hirsch, G.; Kitas, G.; Klocke, R. Intra-Articular Corticosteroid Injection in Osteoarthritis of the Knee and Hip: Factors Predicting Pain Relief—A Systematic Review. *Semin. Arthritis Rheum.* **2013**, *42* (5), 451–473.
117. Thakur, M.; Dawes, J. M.; McMahon, S. B. Genomics of Pain in Osteoarthritis. *Osteoarthr. Cartil.* **2013**, *21* (9), 1374–1382.
118. Bratus, A.; Aeschlimann, A.; Russo, G.; et al. Candidate Gene Approach in Genetic Epidemiological Studies of Osteoarthritis-Related Pain. *Pain* **2014**, *155* (2), 217–221.
119. Palit, S.; Sheaff, R. J.; France, C. R.; et al. Serotonin Transporter Gene (5-HTTLPR) Polymorphisms Are Associated with Emotional Modulation of Pain but Not Emotional Modulation of Spinal Nociception. *Biol. Psychol.* **2011**, *86* (3), 360–369.
120. Hall, K. T.; Loscalzo, J.; Kaptchuk, T. J. Genetics and the Placebo Effect: The Placebome. *Trends Mol. Med.* **2015**, *21* (5), 285–294.
121. Nielsen, L. M.; Olesen, A. E.; Branford, R.; et al. Association between Human Pain-Related Genotypes and Variability in Opioid Analgesia: An Updated Review. *Pain Pract.* **2015**, *15* (6), 580–594.
122. Kraus, V. B.; Stabler, T. V.; Luta, G.; et al. Interpretation of Serum C-Reactive Protein (CRP) Levels for Cardiovascular Disease Risk Is Complicated by Race, Pulmonary Disease, Body Mass Index, Gender, and Osteoarthritis. *Osteoarthr. Cartil.* **2007**, *15* (8), 966–971.
123. Huebner, J. L.; Landerman, L. R.; Somers, T. J.; et al. Exploratory Secondary Analyses of a Cognitive-Behavioral Intervention for Knee Osteoarthritis Demonstrate Reduction in Biomarkers of Adipocyte Inflammation. *Osteoarthr. Cartil.* **2016**, *24* (9), 1528–1534.
124. Nwosu, L. N.; Allen, M.; Wyatt, L.; et al. Pain Prediction by Serum Biomarkers of Bone Turnover in People with Knee

- Osteoarthritis: An Observational Study of TRAcP5b and Cathepsin K in Osteoarthritis. *Osteoarthr. Cartil.* **2017**, *25* (6), 858–865.
125. Neogi, T.; Felson, D.; Niu, J.; et al. Association between Radiographic Features of Knee Osteoarthritis and Pain: Results from Two Cohort Studies. *BMJ* **2009**, *339*, b2844.
126. Collins, J. E.; Losina, E.; Nevitt, M. C.; et al. Semi-Quantitative Imaging Biomarkers of Knee Osteoarthritis Progression: Data from the FNIH OA Biomarkers Consortium. *Arthritis Rheumatol.* **2016**, *68* (10), 2422–2431.
127. Eckstein, F.; Collins, J. E.; Nevitt, M. C.; et al. Brief Report: Cartilage Thickness Change as an Imaging Biomarker of Knee Osteoarthritis Progression: Data from the Foundation for the National Institutes of Health Osteoarthritis Biomarkers Consortium. *Arthritis Rheumatol.* **2015**, *67* (12), 3184–3189.
128. Hunter, D.; Nevitt, M.; Lynch, J.; et al. Longitudinal Validation of Periarticular Bone Area and 3D Shape as Biomarkers for Knee OA Progression? Data from the FNIH OA Biomarkers Consortium. *Ann. Rheum. Dis.* **2016**, *75* (9), 1607–1614.
129. Yusuf, E.; Kortekaas, M. C.; Watt, I.; et al. Do Knee Abnormalities Visualised on MRI Explain Knee Pain in Knee Osteoarthritis? A Systematic Review. *Ann. Rheum. Dis.* **2011**, *70* (1), 60–67.
130. Valdes, A. M.; Doherty, S. A.; Zhang, W.; et al. Inverse Relationship between Preoperative Radiographic Severity and Postoperative Pain in Patients with Osteoarthritis Who Have Undergone Total Joint Arthroplasty. *Semin. Arthritis Rheum.* **2012**, *41* (4), 568–575.
131. Haroutiunian, S.; Nikolajsen, L.; Finnerup, N. B.; et al. The Neuropathic Component in Persistent Postsurgical Pain: A Systematic Literature Review. *Pain* **2013**, *154* (1), 95–102.
132. Wenham, C. Y.; Balamoody, S.; Grainger, A. J.; et al. The Responsiveness of Novel, Dynamic, Contrast-Enhanced Magnetic Resonance Measures of Total Knee Synovitis after Intra-Articular Corticosteroid for Painful Osteoarthritis. *Osteoarthr. Cartil.* **2014**, *22* (10), 1614–1618.
133. Davis, A. J.; Smith, T. O.; Hing, C. B.; et al. Are Bisphosphonates Effective in the Treatment of Osteoarthritis Pain? A Meta-Analysis and Systematic Review. *PLoS One* **2013**, *8* (9), e72714.
134. Siebuhr, A. S.; Bay-Jensen, A. C.; Jordan, J. M.; et al. Inflammation (or Synovitis)-Driven Osteoarthritis: An Opportunity for Personalizing Prognosis and Treatment? *Scand. J. Rheumatol.* **2016**, *45* (2), 87–98.
135. MacGregor, A. J.; Li, Q.; Spector, T. D.; et al. The Genetic Influence on Radiographic Osteoarthritis Is Site Specific at the Hand, Hip and Knee. *Rheumatology (Oxford)* **2009**, *48* (3), 277–280.
136. Haugen, I. K.; Slatkowsky Christensen, B.; Boyesen, P.; et al. Increasing Synovitis and Bone Marrow Lesions Are Associated with Incident Joint Tenderness in Hand Osteoarthritis. *Ann. Rheum. Dis.* **2016**, *75* (4), 702–708.
137. Wang, X.; Blizzard, L.; Jin, X.; et al. Quantitative Assessment of Knee Effusion-Synovitis in Older Adults: Association with Knee Structural Abnormalities. *Arthritis Rheumatol.* **2016**, *68* (4), 837–844.
138. Zhang, Y.; Nevitt, M.; Niu, J.; et al. Fluctuation of Knee Pain and Changes in Bone Marrow Lesions, Effusions, and Synovitis on Magnetic Resonance Imaging. *Arthritis Rheum.* **2011**, *63* (3), 691–699.
139. La Porta, C.; Bura, S. A.; Llorente-Onaindia, J.; et al. Role of the Endocannabinoid System in the Emotional Manifestations of Osteoarthritis Pain. *Pain* **2015**, *156* (10), 2001–2012.
140. Mabey, T.; Taleongpong, P.; Udomsinprasert, W.; et al. Plasma and Synovial Fluid Autotaxin Correlate with Severity in Knee Osteoarthritis. *Clin. Chim. Acta* **2015**, *444*, 72–77.
141. Li, L.; Jiang, B. E. Serum and Synovial Fluid Chemokine Ligand 2/Monocyte Chemoattractant Protein 1 Concentrations Correlates with Symptomatic Severity in Patients with Knee Osteoarthritis. *Ann. Clin. Biochem.* **2015**, *52* (Pt 2), 276–282.
142. Simao, A. P.; Mendonca, V. A.; de Oliveira Almeida, T. M.; et al. Involvement of BDNF in Knee Osteoarthritis: The Relationship with Inflammation and Clinical Parameters. *Rheumatol. Int.* **2014**, *34* (8), 1153–1157.
143. Daghestani, H. N.; Pieper, C. F.; Kraus, V. B. Soluble Macrophage Biomarkers Indicate Inflammatory Phenotypes in Patients with Knee Osteoarthritis. *Arthritis Rheum.* **2015**, *67* (4), 956–965.
144. Dong, T.; Chang, H.; Zhang, F.; et al. Calcitonin Gene-Related Peptide Can Be Selected as a Predictive Biomarker on Progression and Prognosis of Knee Osteoarthritis. *Int. Orthop.* **2015**, *39* (6), 1237–1243.
145. Huo, L. W.; Ye, Y. L.; Wang, G. W.; et al. Fractalkine (CX3CL1): A Biomarker Reflecting Symptomatic Severity in Patients with Knee Osteoarthritis. *J. Investig. Med.* **2015**, *63* (4), 626–631.
146. Dundar, U.; Asik, G.; Ulasli, A. M.; et al. Assessment of Pulsed Electromagnetic Field Therapy with Serum YKL-40 and Ultrasonography in Patients with Knee Osteoarthritis. *Int. J. Rheum. Dis.* **2016**, *19* (3), 287–293.
147. Guan, J.; Liu, Z.; Li, F.; et al. Increased Synovial Fluid YKL-40 Levels Are Linked with Symptomatic Severity in Knee Osteoarthritis Patients. *Clin. Lab.* **2015**, *61* (8), 991–997.
148. Aslam, I.; Perjar, I.; Shi, X. A.; et al. Associations between Biomarkers of Joint Metabolism, Hand Osteoarthritis, and Hand Pain and Function: The Johnston County Osteoarthritis Project. *J. Rheumatol.* **2014**, *41* (5), 938–944.
149. Ke, X.; Jin, G.; Yang, Y.; et al. Synovial Fluid HMGB-1 Levels Are Associated with Osteoarthritis Severity. *Clin. Lab.* **2015**, *61* (7), 809–818.
150. Liu, Y.; Peng, H.; Meng, Z.; et al. Correlation of IL-17 Level in Synovia and Severity of Knee Osteoarthritis. *Med. Sci. Monit.* **2015**, *21*, 1732–1736.